September 12, 2000

Hon. Assistant Commissioner for Patents
Box Patent Appln
Washington, D.C. 20231

Re: New Nonprovisional Continuation, Division or Continuation-In-Part
Patent Application under 37 CFR §1.53(b) in U.S.
Applicant(s): CLASSEN, John Barthelow
Title: METHOD AND COMPOSITION FOR AN EARLY VACCINE TO PROTECT
AGAINST BOTH COMMON INFECTIOUS DISEASES AND CHRONIC
IMMUNE MEDIATED DISORDERS OR THEIR SEQUELAE
Atty's Docket: CLASSEN=1D

Sir:

1. [X] The present nonprovisional application is a [X] Continuation
         [ ] Divisional [ ] Continuation-in-part of prior application No.
         08/591,651. **This application is not a CPA.** The prior
         application is not being abandoned in favor of this application.
    [X] Incorporation By Reference. The entire disclosure of the prior
         application, from which a copy of the oath or declaration is
         supplied herewith, is considered as being part of the disclosure
         of the accompanying application and is hereby incorporated by
         reference therein.

2. Attached herewith is the above-identified nonprovisional application
under 37 CFR §1.53(b) for Letters Patent including:

    [X] Specification (110 pages), claims (30 pages) and abstract (1
         page). The [X] specification [ ] claims and [X] abstract are
         copies of those filed in prior application no. 08/591,651.
    [X] 3 Sheets Drawings (Figures 1-3).
         [ ] Formal   [X] Informal
         [X] Figs. 1-3 of the drawings are copies of those filed in
         prior application no. 08/591,651.
         [ ] Figs. _____ of the drawings are new.
    [X] Declaration and Power of Attorney (1 page)
         [ ] Newly executed [X] Copy from prior application no. 08/591,651
    [ ] Preliminary Amendment
    [ ] Supplemental Preliminary Amendment
    [ ] Information Disclosure Statement with () references
    [X] PTO-1449 or PTO-892 forms from prior application(s)
    [ ] A verified statement to establish small entity status under 37
         CFR §1.9 and 37 CFR §1.27
    [X] A verified statement claiming small entity status was filed on
         February 12, 1996 in prior application no. 08/591,651. Status is
         still proper and desired.
    If no declaration is attached, or the basic filing fee is not paid,
accord this application a filing date, and notify counsel pursuant to 37 CFR
§1.53(f).

[X]   Credit Card Payment Form, PTO-2038, authorizing payment in the amount of $1,753.00 is attahced.

[X]   The filing fee calculated as follows:

CLAIMS AS FILED				
FOR	NUMBER FILED	NUMBER EXTRA	RATE	BASIC FEE $ 690.00
TOTAL CLAIMS	148 - 20=	128	x 22	2,816.00
INDEPENDENT CLAIMS	2 - 3=	0	x 82	0
[ ] Multiple Dependent Claim Presented			x270	
[X] Reduction of 1/2 for small entity				-$1,753.00
			TOTAL FILING FEE	$1,753.00

[ ]   Any additional fee required by the filing of an enclosed preliminary or supplemental preliminary amendment has been calculated as shown below:

	CLAIMS REMAINING AFTER AMENDMENT		HIGHEST NO. PREVIOUSLY PAID FOR	PRESENT EXTRA	RATE	CALCULATION
TOTAL	*	MINUS	**	=	x $ 22.00	$
INDEP	*	MINUS	***	=	x $ 82.00	$
[ ] Multiple Dependent Claim Presented					x $270.00	$
Total of Above Calculations =						$
Reduction by 1/2 for filing by small entity						- $
Total Additional Fee =						$

[ ] _____.

[X]   Return Receipt Postcard (in duplicate)

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3. The following inventor(s) were named in the prior application:

FULL NAME OF INVENTOR	FAMILY NAME	FIRST NAME GIVEN	SECOND NAME GIVEN
	CLASSEN	John	Barthelow
RESIDENCE & CITIZENSHIP	CITY	STATE OR COUNTRY	CITIZEN OF
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FULL NAME OF 2ND INVENTOR	FAMILY NAME	FIRST NAME GIVEN	SECOND NAME GIVEN
RESIDENCE &	CITY	STATE OR COUNTRY	CITIZEN OF
POST OFFICE ADDRESS	POST OFFICE ADDRESS	CITY	ST. & ZIP/COUNTRY
FULL NAME OF 3RD INVENTOR	FAMILY NAME	FIRST NAME GIVEN	SECOND NAME GIVEN
RESIDENCE & CITIZENSHIP	CITY	STATE OR COUNTRY	CITIZEN OF
POST OFFICE ADDRESS	POST OFFICE ADDRESS	CITY	ST. & ZIP/COUNTRY
FULL NAME OF 4TH INVENTOR	FAMILY NAME	FIRST NAME GIVEN	SECOND NAME GIVEN
RESIDENCE & CITIZENSHIP	CITY	STATE OR COUNTRY	CITIZEN OF
POST OFFICE ADDRESS	POST OFFICE ADDRESS	CITY	ST. & ZIP/COUNTRY

(The latest inventor signed §1.63 Declaration in the parent application or its predecessors is hereby incorporated by reference. In the event of discrepancy between the Declaration and the list above, the Declaration controls and the list above is amended accordingly.)

      [XX]   The inventors of the present application are the same as those on the prior application.

      [   ]   Please delete the following inventors who were named on the prior application, but are not inventors of the invention being claimed in the present application:

[ ]   The following additional inventors are to be named on the instant
      application: _____

4.   [ ]   The benefit under 35 USC §119 is claimed of the filing date of:
            Application No. _____ in _____ on _____. A
            certified copy of said priority document [ ] is attached [ ] was
            filed in progenitor case _____ on _____.
            Application No. _____ in _____ on _____. A
            certified copy of said priority document [ ] is attached [ ] was
            filed in progenitor case _____ on _____.

5.   [X]   Amend the specification by inserting before the first line the
            sentence:
            --This is a continuation of copending parent application Serial
            No. 08/591,651, filed February 12, 1996, now pending, which is
            the national stage of PCT/US94/08825, filed August 4, 1994, which
            is a continuation-on-part of 08/104,529, filed August 12, 1993.--

6.   [X]   Certain documents were previously cited or submitted to the
            Patent and Trademark Office in the following prior application
            08/591,651 and 08/104,529, which is relied upon under 35 USC
            §120. Applicants identify these documents by attaching hereto
            one or more form PTO-1449 and/or form PTO-892 listing these
            documents, and request that they be considered and made of record
            in accordance with 37 CFR §1.98(d). Per 37 CFR §1.98(d), copies
            of these documents need not be filed in this application.

            While an IDS filed under §1.97 must contain a "list of all
            patents, publications or other information submitted for
            consideration by the Office", see §1.98(a)(1), the only
            requirement for the list is that it provide the information set
            forth in §1.98(b). There is no requirement that a form PTO-1449
            be used (MPEP §609 merely says that use of this form is
            "encouraged") and no prohibition on submitting a copy of a form
            PTO-1449 or form PTO-892 from a prior case. Indeed, the re-use
            of such forms is desirable as it avoids error in transferring the
            information, and evidences that the reference was considered in
            a prior application. A previously accepted PTO-1449, or an
            examiner-prepared PTO892, necessarily complies with §1.98(b).

7.   If reliance is made on a declaration filed in a prior application, then
      [X]   The undersigned attorney of record hereby revokes the powers of
            attorney of:

            ___G. KEVIN TOWNSEND, Reg. No. 34,033   ___JOHN E. TARCZA, Reg. No. 33,638
            ___JEROME J. NORRIS, Reg. No. 24,696   ___RONALD R. SNIDER, Reg. No. 24,962
            ___TERRELL P. LEWIS, Reg. No. 29,227   ___SANDY LIVNAT, Reg. No. 33,449
            ___BARRY A. EDELBERG, Reg. No. 31,012   ___THERESA BANTA, Reg. No. 34,543
            XX_A. FRED STAROBIN, Reg. No. 18,453   XX_NICK BROMER, Reg. No. 33,478

                                        4

[ ]   The undersigned attorney of record hereby appoints associate
       power of attorney, to prosecute this application and to transact
       all business in the Patent and Trademark Office in connection
       therewith to:
       __IVER P. COOPER, Reg. No. 28,005 __NORMAN J. LATKER, Reg. No. 19,963
       __*ALLEN C. YUN, Reg. No. 37,971
          (* Patent Agent)

8.   [X]   The Commissioner is hereby authorized to charge payment of the
             following additional fees associated with this communication or
             credit any overpayments to Deposit Account No. 02-4035:
             [X] Any additional filing fees required under 37 CFR §1.16.
             [X] Any patent application processing fees under 37 CFR §1.17.

       [X]   The Commissioner is hereby authorized to charge payment of the
             following fees, based on any paper filed during the pendency of
             this application or any CPA thereof, to effect any amendment,
             petition, or other action requested in said paper or credit any
             overpayments to Deposit Account No. 02-4035:
             [X] Any patent application processing fees under 37 CFR §1.17.
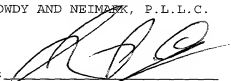   [ ] The issue fee set in 37 CFR §1.18 at or before mailing the
                  Notice of Allowance, pursuant to 37 CFR §1.311(b).
             [X] Any filing fees under 37 CFR §1.16 for presentation of extra
                       claims.
             [X] If a paper is untimely filed in this or any CPA thereof by
                  Applicant(s), the Commissioner is hereby petitioned under 37
                  CFR. §1.136(a) for the minimum extension of time required
                  to make said paper timely. In the event a petition for
                  extension of time is made under the provisions of this
                  paragraph, the Commissioner is hereby requested to charge any
                  fee required under 37 CFR §1.17 to Deposit Account 02-4035.

       [X]   The Commissioner is hereby authorized to credit any overpayment
             of fees accompanying this paper to Deposit Account No. 02-4035.

                          Respectfully submitted,
                          BROWDY AND NEIMARK, P.L.L.C.

                          By:_____
                             Iver P. Cooper
                             Registration No. 28,005

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METHOD AND COMPOSITION FOR AN EARLY VACCINE TO PROTECT AGAINST BOTH COMMON INFECTIOUS DISEASES AND CHRONIC IMMUNE MEDIATED DISORDERS OR THEIR SEQUELAE

BACKGROUND OF THE INVENTION

5   ### Field of the Invention

The present invention involves the fields of immunology and medicine, and more particularly relates to immunization methods, and compositions used therewith, for immunizing young mammals, such as human infants and children, against at least one chronic

10 immune mediated disorder, and, preferably, also against at least one infectious disease.

Related Background Art

Human Pediatric Immunization. A number of severe childhood diseases can strike early in life. Pertussis may pose a serious

15 threat to infants under three months of age, and, during the heyday of the polio epidemic, paralytic cases were reported in the 6-12 month age group with disturbing frequency.

Consequently, to avoid any gap in immunity, it was thought desirable to initiate immunization before infants lost passive

20 protection from maternal antibodies. However, the presence of maternal antibodies can modify or suppress the infant's response to immunization, especially if the vaccine preparations are of low potency. The maturity of the infant's immune system is also a consideration, and premature immunization can result in

25 immunological paralysis.

Accordingly, it has been generally recommended to postpone immunizations in developed countries where maternal antibodies

provide protection against infant infectious diseases until after the age of 2 to 6 months, when the modifying effect of the maternal antibodies had disappeared. Therefore, immunizations should induce an active immune response before the infant loses this passive protection, so that there will be continuous protection from birth without any gap in immunity to natural diseases.

More recently, it has generally been recommended to begin infant immunizations, such as DTP (diphtheria, tetanus and pertussis) and OPV (oral polio virus), at younger ages, and many countries have carried out successful immunization studies and programs beginning at 6-8 weeks of age in developed countries (Expanded Programme on Immunization (1984); (1985)). Accordingly, current recommendations for infant immunization is to provide routine DTP and OPV immunization initiated at no earlier than 6 weeks of age in all developed countries.

The literature on immunization schedules for pediatric vaccines is voluminous, but the examples which follow indicate what has been tried.

Provenzano et al., New England J. Med., 273:959-965 (1965) gave a first dose of a plain pertussis vaccine at 6-24 hours after birth. In Group I, two more such doses were given at three week intervals, and then two doses of combined diphtheria, tetanus and pertussis vaccine (DTP) at one month intervals. In Group II, the initial plain pertussis immunization was followed by three DTP injections at one month intervals.

The authors reported that the immunization response in both groups was inadequate, and attributed it to immunological paralysis "induced by the vigorous immunization schedule employed

and the initiation of immunization on the first day of life."
In view of earlier studies, they recommended that immunization
not be attempted under three weeks of age.

5   Dengrove, et al., Pediatric Res., 20:735 (1986) gave a first
dose of DTP to infants before 4 days of age, and further doses
at 2, 4 and 6 months of age. The immune responses to diphtheria
and tetanus immunogens were acceptable, in contrast to their
previous demonstration that "an early neonatal dose of DTP
resulted in a lowered pertussis antibody response in the subgroup
10 of infants who had low maternally acquired levels of antibody."

     Only 45 infants were treated, and therefore, even if this
schedule could be effective to inhibit diabetes, it is unlikely
that any of the infants actually were benefited (immune-mediated
diabetes occurs in only one out of 200-300 individuals).

15   The immunization protocol used by Baraff, et al.,
Pediatrics, 73: 38-42 (1984) was similar, but the technology used
to evaluate the immune response was more sophisticated. It was
found that the IgG response to the pertussis lymphocytosis -
promoting toxin (LPT) was lower and the IgM anti-FHA (filamentous
20 hemagglutinin) response higher in the early immunization group
than in controls. The authors were of the opinion that the anti-
LPT response was of greater clinical significance and therefore
concluded that neonatal immunization may be disadvantageous.
This would have discouraged use of both Dengrove's and Baraff's
25 protocols.

     Perkins, et al., British Medical J., 68-71 (July 12, 1958)
investigated the response of infants to immunization with a
killed poliomyelitis vaccine. The first dose was given to Group
A at 1 week of age, Group B at six weeks of age, and Group C at

ten weeks of age. A second dose was given four weeks later. Three different virus types were tested in these three groups.

Perkins et al. found that maternally transmitted antibodies interfered with the immune response of the infants to the vaccines. Maternal antibodies declined with age (the half life was about 21 days). The type 2 vaccine was the least susceptible to this interference, but according to table 4, it too, felt it (60% of Group C infants responded, as compared to 35% of those in Group A).

Based on these findings, Perkins et al. concluded "in order to avoid the inhibiting effect of the placentally transmitted antibody, immunization should at present be delayed until six to nine months after birth."

Another study, by Spigland and Goldblum, Pediatrics 25:812-821 (1960) divided infants into groups A (1 and 2 months old), B (3 and 4 months old), and C (5 and 6 months old). Primary immunizations was either at (a) 0 and 21 days, or (b) 0, 7 and 21 days from the first immunization. The vaccine was the formalin-inactivated salk poliomyelitis vaccine. The authors concluded that "presence of maternal antibody seemed to interfere with active production of antibody," and that "the greater the age of primary immunization, the better the response."

In a recent pulse immunization study by John, British Medical Journal 289:88 (1984) the first dose of an _oral_ poliomyelitis vaccine was given at 7, 14, 21, 28, 35 or 42 days of age, and the second and third doses at intervals of four weeks. The immune response to the oral vaccine, unlike the parenteral vaccine discussed previously, did not appear to be affected by the age of the infant. The authors recommended that

children be immunized with the polio vaccine at 1 and 5 weeks, and with polio-plus-DPT at 9, 13, and 17 weeks. The present inventor believes that this immunization schedule would be disadvantageous as the late administration of pertussis would
5 promote the development of diabetes and counteract any anti-diabetic effect of the early polio vaccine dosage.

Barrett, Jr., et al., J. Am. Med. Asso., 167:1103-6 (1958) considered whether it would be advantageous to combine the polio and DPT vaccines. The tetravalent vaccine was administered to
10 children ranging in age from 2 1/2 months to 5 years. Only polio antigen response was measured. The study found that "older children respond much more dramatically than do the infants."

Barrett, Jr. et al., Pediatrics, 30:720 (1962) gave a series of polio-DPT inoculations, beginning at various ages, and then
15 at 1, 2, 3 and 4 months post-initial immunization. The first immunization was at (A) 1-2 days old, (B) 1-2 months old, (C) 3-4 months old or (D) 5-6 months old. Based on their observations, the authors recommended that the initiation of both polio and pertussis immunizations be withheld until the infants was three
20 months of age.

A rather extensive review of the literature on DPT and oral poliomyelitis vaccine (OPV) immunizations has been given by Halsey and Galazky Bull. World Health Org., 63:1151-69 (1985). They compare the antibody response following one dose of OPV at
25 1-12 weeks of life (Table 1) with that to 2-3 doses beginning at 6-8 weeks of life (Table 2), and recommend that in countries where polio-myelitis has not been controlled, trivalent OPV be given at birth and at 6, 10 and 14 weeks of age. Pertussis vaccine schedules are reviewed in Table 3. The response to

immunization beginning at 4 or more weeks was said to be almost as good as the results obtained by beginning at eight or more weeks. They recommended initiating DPT at six weeks of age.

Infants as young as 1-2 months old at the time of initial immunization have received a <u>Hemophilus influenza</u> (bacterial meningitis) (Hibtiter) vaccine (three doses at two month intervals.) Madore, et al., Pediatrics, 85:331-337 (1990). However, Madore, et al. stated that older children responded better than younger children, and their employer, Praxis Biologics, eventually recommended that its Hibtiter vaccine be given to children at 2 to 71 months of age (see PDR). Neonatal vaccination with Bacille Calmette-Guerin (BCG), and its impact on malignant disease, is briefly addressed by Grange and Stanford, Tubercle 71:61-64 (1990). A four dose vaccination schedule has been used to interrupt perinatal vertical transmission of hepatitis B virus, the first being given in the first week of life, and others at 1, 3 and 6 months of age.

While vaccines are subject to safety review, both before and after marketing approval, prior to the present invention, chronic immune mediated diseases, such as diabetes mellitus, were not considered vaccine complications. Many vaccines have been approved based on studies of only a few thousand recipients followed for only a short time after receiving the vaccine. These studies are inadequate to detect the effect of vaccines on chronic immune mediated diseases that are rare and don't develop until many years after a recipient received a vaccine. Current trials with new vaccines only compare results to people who received standard immunization. If both groups received vaccination starting at 2 months, which is now discovered to be

associated with the increased risk then both groups will be associated with a higher than necessary risk of diabetes and other chronic immune mediated disorders. Additionally, current vaccine trials are not designed to look for complications of

5   diabetes or other chronic immune mediated disorders which may not occur until 15 years, or more, after a person is immunized.

Likewise bioassays performed to show safety of vaccines are inadequate because the animals used are from strains that are not susceptible to vaccine induced chronic immune mediated diseases,

10 such as diabetes.

The lack of concern over the ability of vaccines to induce a chronic immune mediated disorder (e.g., but not limited to, diabetes) is further evidenced by the lack of warnings on package inserts and labels of such products about such diseases.

15 As shown above, background art favors administration schedules that induce high levels of antibodies protective against the targeted infectious agent, and therefore typically require beginning immunization at a relatively late age (e.g., for DTP, at two months). The data presented in the present

20 specification shows that such an immunization schedule can cause or at least substantially contribute to the development of chronic immune mediated disorders, e.g. diabetes, as discussed herein.

Animal Pediatric Immunization. Lee, U.S.P. 4,857,318 intra-

25 peritoneally administered a vaccine against Bordetella pertussis to pigs when 7 and 21 days old, or 6 and 12 days old. Intra-peritoneal administration of vaccines to humans is usually

undesirable because of the risk of perforating the intestine. Green, U.S.P. 4,625,015 administered an influenza immunogen to mice at 28, 42 and 56 days after birth. The immunogen was adjuvanted with Freund's complete adjuvant, which is too toxic
5   for human use. In vaccinating humans against influenza, FCA would have to be omitted, with a consequent reduction in the overall immunizing effect. Green does not explain how to adapt his agent or protocol for immunizing humans.

Kuzehara, U.S.P. 5,151,023 administered a single dose of a
10 combined Hepatitis A and B vaccine to guinea pigs and mice, subcutaneously, at 28 days of age.

Chronic Immune Mediated Disorders. Many chronic immune mediated disorders, such as immune mediated cancers and hyperactive immune responses, can be induced or inhibited by
15 cells of the immune system. Environmental stimuli can affect whether those genetically predisposed to a chronic immune mediated disorder will develop symptoms or not. For example, it is not uncommon for one identical twin to have a chronic immune mediated disorder (e.g., type I diabetes mellitus) and the other
20 identical twin to be free of the disorder.

Several methods have been used to modulate an immune response in order to treat autoimmunity or early stages of diabetes mellitus (e.g., as disclosed in U.S. patent No. 4,710,380).
25   Immunosuppressive agents, including both general immuno- suppressants and antigen specific tolerizing agents, have been used to some extent to inhibit or treat chronic immune mediated disorders like autoimmunity. General immunosuppressants can lead

to overwhelming infections and other toxicities as in bone destruction associated with corticosteroids and kidney disease associated with cyclosporine.

Antigen-specific agents that cross-react to a specific autoantigen have been employed to down-regulate immune responses to a particular autoantigen. An example of this approach is contained in PCT patent application (PCT/US90/01397, WO/10449) which discloses the use of antigens which cross-react with a 65Kd heat shock protein as tolerizing agents. Alternatively, PCT/US91/00240 (WO 92/00755) discloses utilizing sub-immunogenic amounts of an antigen which cross-reacts to alloimmune serum, in order to tolerize individuals. The problem with antigen specific agents is that one often does not know all the autoantigens involved in an autoimmune disease and the mechanism requires knowledge of such molecules.

Effect of Immunogens on Immune Disorders. Harada, et al. (1990) reported that a single intravenous injection of live BCG into 5 or 10 week old NOD mice suppressed insulitis and overt diabetes, while an injection into 15 week old mice was somewhat less suppressive.

The dose given by Harada, 0.25-1 mg, is, on a per kg body weight basis (about 8-32 mg/kg) equivalent to a human dose which would not be considered pharmaceutically acceptable. It is not unclear whether Harada's findings regarding the effects of immunication with a live organism are extrapolatable to immunization with inactivated organisms or purified antigens as a live organism which provides continuing exposure of the host to the organism's antigens. BCG's effect may be attributable to its heat shock protein, a known tolerogen, rather than to a

specific immunogenic effect. BCG vaccination has also been inconclusively reported to be associated epidemiologically with both an increased (Salonen, 1975) and decreased (Hems, 1971) incidence of childhood leukemia (Grange, 1990).

Huang, et al (1984) administered one dose of a whole cell _Bordetella pertussis_ vaccine to mice which were at least 45 days old. The dose was given from 10 days before to 30 days after injection of streptozotocin (STZ), which produces a form of insulin-dependent diabetes. The pertussis vaccine aborted the development of IDD as a result of a single injection of STZ. Kolb, et al (1987) also looked at the effects of a pertussis vaccine in a streptozotocin-induced diabetes mouse model. The streptozotocin was administered when the mice were 8-11 weeks of age. The vaccine was given at day -3, +8, or +14 relative to STZ initiation. Given at day -3, it partially suppressed the hyperglycemia, while when given at days -8 and -14, it strongly enhanced it.

As admitted by Huang et al. (1991), "results from streptozotocin-induced IDDM experiments are difficult to extrapolate to type I IDDM because the correlation between chemically-induced diabetes and a "natural" development of autoimmune diabetes is unclear." Consequently, Huang et al (1991) examined the anti-diabetic effect of pertussigen in the genetically predisposed NOD mouse. These mice were give four injections of pertussigen at four week intervals, starting when the mice were 2 (Group 1) or 4 (Group 2 and 3) weeks of age. According to the authors, "although the time at which IDDM was first observed was delayed by several weeks, the incidence rates

were not significantly different from those of untreated control NOD mice."

Toyota et al (1978) administered the islet activating protein of Bordetella pertussis to spontaneously diabetic rats aving a weight of 300-400g. While the age of the rats is not stated, this body weight could not be attributed to a rat younger than 42 days. Furthermore, the animals were already diabetic before the administration of the protein thus the administration was intended as a therapy as opposed to a method of immunization.

It has recently been suggested that vaccination against measles may influence a reduction in the incidence of diabetes, however the data obtained was inconclusive because it failed to look at those who did not receive the vaccine and who did not develop the disease (Blom, 1991). These individuals are here discovered to in fact be at lower risk for developing diabetes than those who received the vaccine.

Effects of Immunomodulators on Immune Disorders. Several papers discuss the effect of OK-432, a streptococcal preparation, on diabetes in mice. (Toyota, et al. 1986; Shintani, et al. 1990). Toyota et al., 1986, gave two clinical units of OK-432 to mice every week from 4-24 weeks of age. Shintani, et al. (1990) also tested schedules of weekly immunization at 4-15, 4-9, and 10-15 weeks of age. Weekly injections were needed at a dose of approximately 0.1 mg/20g, (5 mg/kg or 100 KE/kg) mouse to provide the protective effect (Shintani, Satoh, Seino, et al 1990), while pharmaceutically acceptable clinical doses would be 0.07 KE/kg or 1400 times less.

OK-432 is a pyretic agent. See Shinoda, et al. Acta Urologica Japonica, 38:1299-13ct (1992); Luh et al., Cancer, 69:674-9 (1992), Imamura, et al., Cancer, 68:259-63 (1991); Ogita, et al., J. Pediatric Surgery, 26:263-8 (1991). While it may be reasonable to prescribe it to a patient with cancer, it would not be clinically indicated for prevention of diabetes in youngsters. Young children are particularly prone to seizures as a result of high fevers. Ogita et al. used OK-432 only to treat surgically unresectable lymphangiomas in children.

Oldstone (1988) inoculated NOD mice with live, unattenuated, lymphocytic choriomeningitis virus (LCMV) either intracerebrally at birth with LCMV ARM536 or intravenously with "clone 13" at six weeks of age. Both treatments prevented the spontaneous development of type 1 diabetes characteristic of NOD mice. This work was elaborated upon by Oldstone, et al. (1990), who inoculated NOD mice intracerebrally, within the first eighteen hours of life, with various LCMV strains and reassortants. Schwimmbeck, et al. (1989) found that acute infection with LCMV at 30 days of age abrogated diabetes in BB rats.

However, these inoculations did not immunize the rodents against LCMV, and therefore did not offer protection against an infectious disease. Indeed, the administration infected the rodents with the virus, which persisted throughout the animal's lifespan, and altered the cytotoxicity of the host's T lymphocytes. The present invention does not contemplate inoculating a subject with a live, unattenuated virus. Lympho-tropic viruses are potentially very dangerous, as the case of HIV illustrates, and it would be difficult to win clinical acceptance of such a virus as a human therapeutic agent. Moreover, intra-

cerebral administration of vaccines to humans is undesirable because of the risks of damaging brain cells or inducing meningitis. In any event, the administration of other viruses has been shown to increase the incidence of diabetes (Guberski

5   et al, 1991: Kilham's rat virus in BB/Wor rats).

Several other agents have to be used in attempts to alter the development of immune mediated disorders. Specific lympho-kines like IL-2 (e.g., according to Serreze et al, 1989) and tumor necrosis factor (e.g., according to Satoh et al, 1989) have

10 also been employed to attempt to treat or prevent immune mediated disorders thought to be caused by lymphokine defects, by using multiple injections. However, purified lymphokines are relatively toxic, and expensive, and have short half lives.

Freund's adjuvant (Sadelain et al, 1990) has also been used

15 to delay the development of diabetes in NOD mice. However, this adjuvant is not suitable for vaccine use in mammals at levels comparable to those used by Sadelein et al, (1990) due to its toxic effects. Freund's adjuvant can cause plasmacytomas as well as granulomas.

20 The use of anti-receptor and immune modulating agents (such as products that can block specific receptors, activate specific receptors, or cause the release of suppressor factors) generally requires repeated injections at high doses, e.g., in the mg/kg of body weight range. Examples of anti-receptor immunogens

25 include monoclonal antibodies (e.g., U.S. Patent No. 4,695,459) and lectins like Concanavalin A (e.g., Pearce and Peterson, 1991). Pharmaceutical use of monoclonal antibodies reactive to the antigen specific T-cell receptor have been shown to be associated with an increase in human lymphoid tumors and thus

anti-receptor ligands reactive to the this T-cell receptor are unlikely to be pharmaceutically acceptable.

In conclusion, it has heretofore not been clearly shown or recognized that pharmaceutically acceptable doses of pharmaceutically acceptable immunogens can prevent chronic immune mediated disorders. The related art has also not demonstrated whether such agents would be of value in preventing chronic immune mediated disorders in mammals, especially humans, that are immunized early in life. The related art has also not shown when or how to administer such agents to a mammal which being immunized early in life against infectious diseases, in order to prevent or reduce the prevalence/incidence/frequency/severity of chronic immune mediated disorders.

Citation of documents herein is not intended as an admission that any of the documents cited herein is pertinent prior art, or an admission that the cited documents is considered material to the patentability of any of the claims of the present application. All statements as to the date or representation as to the contents of these documents is based on the information available to the applicant and does not constitute any admission as to the correctness of the dates or contents of these documents.

SUMMARY OF THE INVENTION

The present invention relates to the discovery that when one or more immunogens, in a pharmaceutically acceptable composition, is first administered at an early age (typically prior to 42 days of age), it can substantially decrease the incidence, frequency, prevalence or severity of, or prevent, at least one chronic immune mediated disorder, and/or a surrogate marker thereof. Usually, at least one of these early administered immunogens is administered with the additional purpose of immunization against an infectious disease. Indeed, the immunogens which thus combat a chronic immune mediated disorder may collectively contribute to protection of the subject against two, three, or even more, infectious diseases.

Without intending to be bound by any theory, early administration of immunogens can cause the release of lymphokines that may accelerate the maturation of the immune system in a manner which reduces the likelihood of development of a chronic immune mediated disorder. The immunization may act in several ways including:

A.   Enhancing destruction of autoimmune prone cells in the thymus;

B.   Enhancing the flow of normal T-cells from the thymus;

C.   Causing peripheral elimination of autoreactive T-cells that have escaped the thymus;

D.   Causing the release of interferons which prevent infection with autoimmune causing viruses; and/or

E.   Causing migration of macrophages into the area of administration as in an injection site and away from an vital organ like the islet cells of the pancreas.   The invading

macrophages have the ability to act as antigen presenting cells and induce an autoimmune response against the vital tissue.

The present invention may therefore be said to relate to early immunization programs which achieves one or more of these activities.

In contrast, the late administration of an immunogen can cause the release of lymphokines which may act as growth factors enabling autoimmune inducing cells to grow.

In preferred embodiments, the immunization schedules of the present invention may include employing initiating immunization prior to 28 days, supraimmunogenic doses, multiple doses prior to 56 or 112 days of age and/or dosing intervals of less than 28 days.

In some embodiments, the immunization schedule comprises administration of at least one non-pediatric immunogen (e.g. anthrax or plague immunogens).

New pharmaceutical agents of the present invention are described that contain both non-pediatric and pediatric immunogens. Methods of manufacturing such agents are also included. Kits are described containing in receptacles at least one non-pediatric immunogen and at least one pediatric immunogen. However, such agents and kits are not required for the practice of the invention.

Methods are also provided for screening immunogenic agents for their ability to modulate the development of at least one chronic immune mediated disorder in a mammal such as a human.

The present invention is especially useful in preventing diabetes mellitus and systemic lupus erythrematosis.

Other objects, features and advantages of the present invention will be clear to those skilled in one or more of the relevant arts, based on the teachings and guidance presented herein.

BRIEF DESCRIPTION OF THE DRAWINGS

In Figures 1-3, the trendlines plot the % diabetic (ordinate) against age (in weeks after birth, abscissa).

Figure 1 depicts data from Example 1. Results show that mice receiving anthrax vaccine (A exp1) reached a peak cumulative incidence of diabetes of 42.1% at 24 weeks of life and no new cases of diabetes were detected through week 36 of life. The control receiving PBS (PBS exp1) showed a steady increase in the cumulative incidence of diabetes and 75% of mice had become diabetic by 36 weeks of life. Those animals receiving the plague vaccine (Plague exp1) had a lower incidence of diabetes than the controls throughout most of their life span. The plague vaccine treated animals had a cumulative incidence of diabetes of 57.9% at 28 weeks and development of diabetes appeared to be leveling off after 26 weeks. Plague vaccine treated animals were sacrificed at 28 weeks.

Figure 2 depicts data from Example 2. Animals were given 1 of 6 treatments: anthrax and the whole cell DTP vaccine (ADPT exp2); anthrax and the diphtheria tetanus vaccine (ADT exp2); anthrax vaccine alone (A exp2); a single injection of DTP vaccine at week 8 (control + DPT exp2); anthrax and the diphtheria tetanus vaccine with the whole cell DTP vaccine substituted for the diphtheria tetanus vaccine at 8 weeks (ADT + DPT exp2); and PBS control (PBS exp2). Data show a single injection of DTP does not prevent diabetes and actually increases the incidence of diabetes as seen in the difference between the group receiving ADT+DTP and ADT. All of the animals in the group receiving ADTP did not develop diabetes.

Figure 3 depicts data from Example 4. BB rats received either DTP and Anthrax vaccines (squares) or nothing (control; stars). Results show at 16 weeks of age 54% of the untreated rats had developed diabetes and or died compared to 20% in the vaccinated group. At 20 weeks of age 54% of the untreated rats had developed diabetes and or died compared to 25% in the vaccinated group. At 32 weeks the results were 54% versus 35% respectively which represents a 34% reduction in the incidence of diabetes. The difference between the two groups were statistically significant at 20 weeks (P=0.027).

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

OF THE INVENTION

Thus, it has now been discovered that the occurrence and/or severity of chronic immune mediated disorders may be substantially prevented, or their symptoms, prevalence, incidence, severity, or frequency reduced, by administration to a mammal, such as a human, of at least one pharmaceutically acceptable dose of a pharmaceutically acceptable immunogenic agent, wherein the first pharmaceutically acceptable dose is given prior to the first 6-8 weeks of life.

It was also unexpectedly found that immunization with conventional pediatric vaccines beginning at 6-8 weeks, which is the common immunization protocol in developed countries, actually can increase the probability that a mammal will develop a chronic immune mediated disorder. Current vaccination protocols, such as childhood active immunization protocols (e.g., as presented in THE MERCK MANUAL, 16th ed., 60-64 and 1944-47 (1992) entirely incorporated herein by reference) may at best only slow the progression of immune mediated disorders, but not prevent them.

While typically the subject of immunization will be a human being, it may also be a nonhuman mammal, especially mammals of the orders Primata (monkeys or apes), Carnivora (especially one of the domesticated species such as dogs and cats), Artiodactyla (especially one of the domesticated species such as horses, goats, pigs, sheep, cows), or Rodenta (especially domesticated or laboratory species such as mice, rats, rabbits and hamsters).

The subject may be healthy at the time of immunization, or may be one suffering from an adverse clinical condition such as prematureness, diarrhea, infection, or fever.

Chronic Immune-Mediated Disorders

A chronic immune mediated disorder is one which lasts longer than two months, but does not include permanent sequela of acute immune response diseases such as seizures and anaphylaxis, nor do such disorders include diseases associated with live virus infections as in subacute sclerosing panencephalitis induced by measles vaccine.   Chronic immune mediated disorders does not include sequela caused by chronic infections by live vaccines. The invention is especially useful in preventing chronic immune disorder which develop at least one year after a vaccination. Thus, an illness like Guillean-Barrea syndrome is not routinely considered an chronic immune mediated disorder.

A growing number of human diseases have been classified as autoimmune in nature (see, Theofilopoulos, A., In: D.P. Stites, et al., eds., Basic and Clinical Immunology, Lange Medical Publications, Los Altos, CA, 1988; and Berkow, supra), which references are entirely incorporated by reference, and the present invention is intended to include as a chronic immune mediated disorder any and all of such diseases, with the exception listed herein, in mammals including humans.

Methods and compositions of the present invention may be used for preventing and/or inhibiting chronic immune mediated disorders including immune mediated cancers and hyperactive immune responses.   Such immune mediated cancers may include lymphoreticular neoplasia, lymphoblastic leukemia, brain tumors, gastric tumors, plasmacytomas, multiple myeloma, leukemia, connective tissue tumors, solid tumors and lymphomas.   Such hyperactive immune responses may include asthma/allergies and autoimmune diseases.   Such allergies may include hay fever,

atopic dermatitis, urticaria, perennial rhinitis, allergic conjunctivitis, pulmonary diseases, food allergies, skin allergies, anaphylaxis (e.g., associated upon exposure to blood products) and pollinosis. Such autoimmune diseases may include

5   type 1 diabetes, conventional organ specific autoimmunity, neurological disease, rheumatic diseases/connective tissue disease, autoimmune cytopenias, and related autoimmune diseases. Such conventional organ specific autoimmunity may include thyroiditis (Graves+Hashimoto's), gastritis, adrenalitis

10 (Addison's), ovaritis, primary biliary cirrhosis, myasthenia gravis, gonadal failure, hypoparathyroidism, alopecia, malabsorption syndrome, pernicious anemia, hepatitis, anti-receptor antibody diseases and vitiligo. Such neurological diseases may include schizophrenia, Alzheimer's disease,

15 depression, hypopituitarism, diabetes insipidus, sicca syndrome and multiple sclerosis. Such rheumatic diseases/connective tissue diseases may include rheumatoid arthritis, systemic lupus erythematous (SLE) or Lupus, scleroderma, polymyositis, inflammatory bowel disease, dermatomyositis, ulcerative colitis,

20 Crohn's disease, vasculitis, psoriatic arthritis, exfoliative psoriatic dermatitis, pemphigus vulgaris, Sjorgren's syndrome. Other autoimmune related diseases may include autoimmune uvoretinitis, glomerulonephritis, post myocardial infarction cardiotomy syndrome, pulmonary hemosiderosis, amyloidosis,

25 sarcoidosis, aphthous stomatitis, and other immune related diseases, as presented herein and known in the related arts. See, e.g., Berkow et al., eds, *supra*, pages 303-364, 710-718, 1083, 1269, 1305-1377, 1338 1677-1684, and 2435-2438 which is entirely incorporated herein by reference.

Type I diabetes mellitus is defined herein as an naturally occurring or spontaneously developing disease of the pancreatic islet cells that is not intentionally induced. This definition is intended to differentiate the natural disease from altered insulin secretion following the deliberate or intentional destruction of pancreatic islet cells through the use of toxic agents, surgery or other bodily insults.

Type II diabetes mellitus will benefit from the invention as a non-limiting example. The etiology of this disorder is poorly understood. However, scientists believe some patients are unable to secrete enough insulin or are resistant to insulin. Scientists have not ruled out that many of these patients fail to produce enough insulin because they have insufficient islet cells. Scientific evidence suggests that failure to produce sufficient insulin in some type II diabetics is due to destruction of islet cells (Niskanen et al 1991). This evidence includes the presence of autoantibodies to islet cells. These patients differ from type I patients in that they produce enough insulin to prevent them from becoming ketotic and they develop disease later in life. A person who is resistant to insulin and needs more insulin to survive would be at a disadvantage if that person had lost a substantial amount of his or her islet cells do to an immune mediated attack earlier in life. A vaccine to protect islet cells would thus be advantages to people pre-disposed to type II diabetes. Accordingly, methods and immuno-genic agents of the present invention provide a treatment for reducing the severity or incidence of diabetes.

There are multiple sequelae to chronic immune mediated disorders. As non-limiting examples, autoimmunity can result in

end organ failure or cancer. Chronic inflammation, as occurs in chronic immune mediated disorders, can cause the release of molecules like serum amyloid which can cause pathology. Serum amyloid is associated with amyloidosis peripherally and senile dementia in the central nervous system.

The present invention is intended to utilize low or non toxic agents that can be used to prevent disease in asymptomatic mammals without the need to screen them for risk of developing chronic immune mediated disorders, however the invention may be used in certain subpopulations at higher risk for developing the disorders than others. Alternatively the invention may be administered to a large number of mammals with different risks for developing a given chronic immune disorder and only certain subpopulations may be shown to benefit statistically from the administration. The subpopulations may include mammals at higher risk than the general population. Non-limiting examples of the subpopulations include those with family history of at least one chronic immune mediator disorder, those who are deemed at high risk because of genetic or biochemical screening of themselves or biological relatives, and those at risk because of an abnormal birth as in premature or small size.

Immunization Schedule

An immunization schedule is a program for the administration of one or more specified doses of one or more specified immunogens, by one or more specified routes of administration, at one or more specified ages of the immunization subject. A supplemental immunization schedule is one intended to supplement

a standard immunization schedule which is commonly followed in the region in which the subject resides.

The immunization schedules of the present invention induce an immune response in the subject sufficient to reduce at least one measure selected from the group consisting of incidence, prevalence, frequency and severity of at least one chronic immune mediated disorder, and/or at least one surrogate marker of the disorder, in a population and/or subpopulation of the mammals. A supplemental immunization schedule is one which has this effect relative to the standard schedule which it supplements. The supplemental schedule may call for additional administrations and/or supraimmunogenic doses of immunogens found in the standard schedule, or for the administration of immunogens not part of the standard schedule. The latter may be non-pediatric immunogens. The full immunization schedules of the present invention may comprise both a standard schedule and a supplemental schedule.

In the method of the present invention, the first administration of an immunogen, especially one which also serves to contribute to immunization against an infectious disease, usually occurs when the subject is less than 42 days of age (in the case of a non-pediatric immunogen, the first administration could be as late as 55 days of age), more preferably less than 28 days of age, still more preferably less than 14 days of age, and most preferably at about 7 days of age. It is permissible to begin the program at an even earlier age, e.g., within the first 24 hours after birth. The "age", it should be noted, is measured from birth and not from conception.

While it is possible that the purposes of the present invention can be served with a single administration, especially

when the immunogen is a strong one, and in that case a single
dosing schedule is within the compass of the present invention,
it is desirable to administer two or more dosings for greater
surety.   Preferably, the number of dosings is at least three,
5   more preferably at least four, and still more preferably, at
least five.   There is no set maximum number of vaccinations,
however it is good clinical practice not to immunize more often
than necessary to achieve the desired effect. (For the purpose
of the appended claims, the administration of two different
10   immunogens, or of two packets of the same immunogen, within a
period of less than 24 hours, is considered a single dosing.)

Example 1, as non-limiting example, shows that just three
vaccinations with a single mild agent, anthrax vaccine, can cause
a profound reduction in diabetes.   Increasing the number of
15   vaccinations increases the effect of a mild agent like the
anthrax vaccine, as seen by the improvement from Example 1 to
Example 2.

When an immunization schedule calls for two or more separate
dosings,   it is necessary to consider the intervals between
20   dosings. The interval between two successive dosings may be the
same throughout the program, or it may change as the subject
ages.   As immunizations are usually incorporated into routine
office checkups, and these usually become less frequent as the
child's health becomes a known quantity, it is not unusual for
25   the interval to lengthen with age.   While the interval can be
shortened, there would have to be a tangible benefit for this to
be acceptable to clinicians.

Preferably, in the immunization schedules of the present
invention, at least during the first 175 days from birth, the

shortest interval between successive dosings is less than 28 days, more preferably, less than 14 days, and still more preferably, about 7 days. Preferably, at least during the first 175 days from birth, the longest interval between successive dosings is less than 28 days, more preferably, less than or about 14 days.

The timing of the initial dosing and the intervals between successive dosings together determine how many dosings occur within a particular period in the subject's life. Within the first 112 days after birth, the subject preferably receives at least three, more preferably at least four, and still more preferably at least five dosings. Moreover, within the first 42 days after birth, the subject preferably receives at least two, and more preferably at least three, dosings.

The immune systems of mice and men mature at comparable rates, with both species capable of mounting immune responses to vaccine antigens by the time the recipients are several months old. A comparison of the experimental and epidemiological examples in this specification supports this conclusion. Subtle differences in the rates of development of the immune systems of mice and humans may be detected however using a broad range of assays including in vivo assays, in vitro assays, in vitro assays and phenotypci cell assays.

In vivo assays include the non-limiting examples of antibody responses and delayed type hypersensitivity responses. The anti-body responses primarily measures B-cell function as well as B-cell T-cell interactions while the delayed type hypersensitivity responses measure T-cell immunity. One may compare antibody titers in the blood following an antigenic challenge. These

levels may be quantitated according to the type of antibody as in the non-limiting examples of IgG, IgG1, IgG2, IgM, or IgD. Once can also compare development of immune systems by determining levels levels of anitbodies and lymphocytes in the blood without antigenic stimulation.

In vitro assays may include the non-limiting examples of determining the ability of cells to divide, provide help for other cells to divide, release lymophokines and other factors, express markers of activation, and lyse target cells. Lymphocytes in mice and man can be compared in in vitro assays. Ideally one wants to compare the lymphocytes from similar sources as in the non-limiting examples of peripheral blood cells, spleenocytes, or lymphnode cells. It is possible however to compare lymphocytes from different sources as in the non-limiting example of peripheral blood cells in humans and splenocytes in mice. Cells may be puprified or left in their natural state as in the non-limiting example of purified B-cells, T-cells, and macrophages as compared to unpurified splenocytes or lymph node cells. Purification may be by any method that gives the desired results. The cells may be tested in vitro for their ability to proliferate using mitogens or specific antigens. Mitogens can specifically test the ability of either T-cells to divide as in the non-limiting examples of concanavalin A and T-cell receptor antibodies, or B-cells to divide as in the non-limiting example of phytohemagglutinin. The ability of cells to divide in the presence of specific anitgens can be determined using a mixed lymphocyte reaction, MLR, assay. Supernatant from the cultured cells may be tested to quantitate the abilty of the cells to secrete specific lymphokines. The cells can be removed from

culture and tested for their ability to express activation antigens. This may be done by any method that is suitable as in the non-limiting example of using antibodies or ligands to which bind the activation antigen as well as probes that bind the RNA coding for the activation antigen.

Phenotypic cell assays can be performed to determine the frequency of certain cell types. Peripheral blood cell counts may be performed to determine the number of lymphocytes or macrophages in the blood. Antibodies may be used to screen peripheral blood lymphocytes to determine the percent of cells expressing a certain antigen as in the non-limiting example of determining CD4 cell counts and CD4/CD8 ratios.

The present invention therefore can include administration of the immunogens to humans when said humans' immune systems are in a state of maturation and responsiveness comparable to that of mice or rats at the times indicated above, in such circumstances as it would be less effective to administer those immunogens to humans at the same chronological ages as they were administered to mice or rats.

Four non-limiting examples of vaccination schedules taught in the specifications are shown in **Table V**. These vaccine schedules were derived from information supplied in the specifications. Detailed descriptions of screening methods discussed herein allow one to develop alterations to the sample immunization schedules without undue experimentation.

Schedule 1 depicts a very intensive immunization schedule where the recipient is immunized in the first week of life and receives subsequent immunizations every two weeks. The recipient receives at least one non-pediatric immuogen along with common

pediatric vaccines. All vaccines are administered during each
scheduled imunization. Additional vaccines may be added to the
vaccination schedule as in the non-limiting example of a
hepatitis A vaccine. Vaccines may be omitted from the vaccina-
tion schedule as in the non-limiting example of the Hemophilus
influenza vaccine. Vaccines may be continued for different
durations of time.

Schedule 2 depicts another possible immunization schedule.
Immunization is started on the second week. The immunization
protocol is less intense since a non-pediatric vaccine is not
administered. An oral polio vaccine is administered instead of
the inactivated vaccine used in Schedule 1. As in Schedule 1,
vaccines may be added or omitted or continued for different
durations of time.

Schedule 3 depicts a third possible immunization schedule.
Vaccines are administered starting soon after birth and
immunization continues every third week. Vaccines are continued
for different durations of time. The non-pediatric vaccine is
administered four times, the hepatitis B vaccine is administered
five times and other vaccines are administered seven times.
Vaccines may be added or omitted or continued for different
durations of time.

Schedule 4 depicts a fourth possible immunization schedule.
Vaccines are started at different times in this schedule. The
Pertussis vaccine and the MMR vaccines are started on week two
while the other vaccines are started soon after birth. Immuniza-
tions are initially administered every two weeks but immunization
frequency is shifted to every third week. Vaccines may be added
or omitted or continued for different durations of time.

It will be appreciated by those skilled in the art that a variety of possible combinations and subcombinations of the various preferred conditions of timing of the first administration, shortest interval, largest interval and total number of administrations (in absolute terms, or within a stated period) exist, and all of these combinations and subcombinations should be considered to be within the inventor's contemplation though not explicitly enumerated here.

The inventor appreciates that it is conceivable that a prior experimenter has, without recognition of its anti-chronic immune-mediated disorder activity, proposed or even practiced an immunization schedule which falls within the present disclosure. If, under the applicable law, such a proposal or practice would be deemed to anticipate or render obvious an invention here claimed, then it is within the inventor's contemplation to excise from the invention the specific embodiment in question, preserving to the maximum degree permitted by law the scope of protection originally sought.

The ability of an immunization program to substantially reduce the incidence or severity of an immune-mediated disorder is preferably determined relative to a standard immunization protocol. The following three schedules are preferred standards:

32

Schedule

Week	1	2	3
0	x	x	HepB
2	x	x	x
4	x	x	HepB
6	DTP,Polio,HepB,HiB	x	x
8	x	DTP,Polio,HepB,HiB	DTP,Polio,HepB,HiB
10	DTP,Polio,HepB,HiB	x	x
12	x	DTP,Polio,HepB,HiB	DTP,Polio,HiB
14	DTP,Polio,HepB,HiB	x	x
16	x	DTP,Polio,HepB,HiB	DTP,Polio,HiB
60	MMR	MMR	MMR

MMR: Measles, Mumps, Rubella
HepB: Hepatitis B
HiB: Hemophilus Influenza
x: No administration

Preferably, a statistically significant ($p<=.05$) reduction of at least about 10% is seen, relative to control employing one of these three schedules or relative to untreated controls. More preferably, the reduction is greater, i.e., at least about 20% or at least about 50%.

When reduction in a surrogate marker is the criterion, the surrogate marker may be selected from the group consisting of an autoantibody, an autoreactive lymphocyte, a biochemical marker of tissue destruction, an biochemical marker of abnormal secretion from an internal body organ, an positive skin reaction to an immunological challenge which indicates immunological hypersensitivity, and an elevated level of antibodies that correlate with hypersensitivity. The abnormal secretion from an internal body organ may be abnormal insulin secretion.

Preferably, one or more of the immunogens administered according to the immunization schedule is given, not only to achieve a beneficial effect with regard to chronic immune-mediated disorders, but also to immunize the subject against an

infectious disease. These infectious diseases include, but are
not limited to, diphtheria, tetanus, pertussis, polio, hepatitis
B, hemophilus influenza, measles, mumps, and rubella.

While the immunogens which can be administered according to
the present invention are discussed in detail in a later section,
certain prefatory remarks regarding their place in the
immunization schedule are appropriate here. The immunization
schedule may provide for one or more administrations of a single
immunogen, multiple immunogens which collectively immunize
against the same or different strains for the same infectious
disease, or multiple immunogens which collectively immunize
against a plurality of different infectious diseases. The
administered immunogens may all be "pediatric immunogens", as
hereafter defined, all "non-pediatric immunogens", or a combina-
tion of both. The immunogens may be separately or simultaneously
administered, and, in the latter case, may be combined into a
single pharmaceutical composition for ease of administration.

Immunogens

Immunogens correspond to a class of molecules that elicit
an immune response through classical immunologic pathways as in
the non-limiting example of the incorporation in an MHC molecule
of an antigen processing cell where the immunogens can
potentially interact with antigen specific T cell receptors.
Alternatively, as another non limiting example, immunogens can
bind to antigen specific binding regions of immunoglobulins which
may lead to stimulating the B lymphocytes (if on the surface of
B lymphocytes), but alternatively could elicit an immune response

through other means, e.g., by the activation of complement, or the modulation of Fc receptors.

Immunogens may be derived from a virus, bacteria, yeast, mold, plant, insect, allogeneic or xenogeneic animal or a

5   molecule, compound or composition that immunologically cross reacts with a naturally occurring immunogen. Such agents may be made from the killed or live bacteria, killed or live viruses, recombinant or chemically synthesized or purified immunogenic agents including antigens, fragments or cross reacting synthetic

10 or recombinantly produced peptides, carbohydrates, lipids or any combination thereof. Such agents can be combined with each other and with vaccines against infectious diseases to substantially prevent or reduce the incidence of immunologic disorders according to the present invention.

15 The term "immunologically cross reacts" refers to molecules that induce antibodies or T-cells that bind to the cross reactive molecule or fragments thereof.

Weak immunogens may be limited to the ability to invoke changes in such immune mediator cells, such as lymphocytes (B

20 and/or T cells), macrophages and natural killer cells, such as the release of lymphokines, altered cell movement, or altered composition of cell surface receptors. Strong immunogens have the additional ability to cause either an humoral immune response (such as, e.g., antibodies to said agent) or a cellular immune

25 response (such as, e.g., a delayed type skin reaction to said agent).

There are several examples of conventional immunogens. The classical example is that of vaccines as in human vaccines. Such vaccines may be classified as living where such agents may

multiply or perform homeostatic metabolic activity in the recipient, as in the live oral polio, live BCG, and live small pox vaccines, as non-limiting examples. Alternatively, conventional vaccines can be classified as inactivated (killed), where

5 such agents have lost their ability to multiply or maintain homeostatic metabolic activity. Non-limiting examples of such killed vaccines include tetanus toxoid, diphtheria toxoid, and the killed whole cell pertussis vaccine. Other non-limiting examples of conventional non-living immunogens are haptens, anti-

10 idiotype antibodies, and nucleic acid molecules, such as DNA or RNA, that can be expressed in cells as immunogenic molecules encoded by such nucleic acids. Alternatively, conventional immunogens may be classified according to their functional or structural properties in a microorganism such as capsular,

15 fimbriae, nuclear, cell wall, membrane, and cytoplasmic immunogens.

Chemically speaking, immunogens of biological origin are most often peptides (including proteins), carbohydrates, glycopeptides, lipids, glycolipids, or lipopeptides.

20 Immunogens of the present invention may be pediatric or non-pediatric immunogens. The term "pediatric immunogens" refers to immunogens that after birth were routinely administered to children less than 16 weeks old, in modern developed nations of moderate latitudes in 1992. These agents include but are not

25 limited to BCG, measles, mumps, rubella, diphtheria, pertussis, hemophilus influenza, tetanus, hepatitis B, and polio.

Non-pediatric immunogens are immunogens not routinely administered to children prior to 112 days in modern developed nations of moderate latitude in 1992, and may include, but are

not limited to, the group consisting of anthrax, plague, encephalitis, meningococcal, meningitis, pneumococcus, pneumonia, typhus, typhoid fever, streptococcus, staphylococcus, neisseria, lyme disease, cholera, E. coli, shigella, leishmania, leprosy,

5   cytomegalovirus (CMV), respiratory syncytial virus, Epstein Barr virus, herpes, influenza, parainfluenza, rotavirus, adenovirus, human immunodeficiency virus (HIV), hepatitis A, NonA NonB hepatitis, varicella, rabies, yellow fever, rabies, Japanese encephalitis, flavivirus, dengue toxoplasmosis, coccidiomycosis,

10 schistosomiasis, and malaria immunogens.

It may be advantageous to include in the immunization schedule at least one immunogen against which the subject is unlikely to possess maternal antibodies; such an immunogen will normally elicit a stronger response and is less likely to cause

15 immunological paralysis.

Immunosuppressants like corticosteroids, azathioprine, cyclosporine, and FK-506 do not activate immune mediator cells and are not considered immunogens in this invention. Tolerogens also are not generally considered immunogens in this invention,

20 except as a tolerogen-immunogen, as described herein. A tolerogen is generally defined as an agent which induces a state of antigen specific immunological unresponsiveness to an antigen that immunologically cross reacts to the agent. Tolerogens are further considered agents that inactivate immune mediator cells

25 like B and T lymphocytes by reacting to their antigen specific binding sight and inactivating the cells in an antigen specific manor. However, if a tolerogen has a component that is clearly immunogenic and causes activation of immune mediator cells resulting in antibody formation or T cell immune responses, then

it can be both a tolerogen and an immunogen. A tolerogen-immunogen in the latter case may be employed in this invention to prevent chronic immune disorders by down regulating cells that do not directly bind to the tolerogen and/or prevent chronic

5 immune responses against organs/antigens that do not cross react immunologically to said tolerogen.

Immunogens are distinct from immune modulators. There are several classes of immune modulators. One class is "immunocyte receptor ligands." Members of this class of agents bind to cell

10 receptors of immune mediator cells in a non-antigen specific manner to cause the induction of an immune response, e.g., as defined herein. One subclass of this group is cytokines. Cytokines that are produced by lymphocytes are termed lympho-kines, whereas peptides produced by monocytes or macrophages are

15 given the term monokines. Thus, the terms cytokines, lympho-kines, and interleukins may be used interchangeably to designate those peptide molecules that modulate host responses to foreign antigens or host injury by regulating the growth, mobility and differentiation of leukocytes and other cells.

20 Known cytokines include interleukins (IL) IL-1 (also endogenous pyrogen (EP), lymphocyte activating factor (LAF), mononuclear cell factor, catabolin, osteoclast activating factor and hematopoetin 1), IL-2 (also T cell growth factor (TCGF)), IL-3 (multicolony stimulating factor (M-CSF), P-cell stimulating

25 factor, WEHI-3B factor, mast-cell growth factor and histamine-producing factor), IL-4 (B-cell growth factor (BCGF), B-cell stimulatory factor-1 (BSF-1), IL-5 (T-cell replacing factor (TRF), B-cell growth factor II (BCGF-II), eosinophil differen-tiation factor (EDF), IL-6 (β_2 interferon (IFN-β_2), B-cell

stimulating factor 2 (BSF-2), 26-kDa protein, hybridoma/plasmacy-
toma growth factor (HPGF or IL-HP-2), hepatocyte stimulating
factor (HSF), and T-cell activating factor (TAF)), IL-7, IL-8
(neutrophil activating protein 1 (NAP-1), IL-10 (also cytokine

5   synthesis inhibitory factor (CSIF); tissue necrosis factors (TNF)
TNFα (also lymphotoxin (LT) and TNFβ (also macrophage derived
TNF); interferons (IFN) IFNα and IFNβ (also type I IFN) and IFNγ
(also type II IFN) and tissue growth factor (TGF) β.

Granulocyte-Macrophage Colony Stimulating Factor or GM-CSF
10   is another non-limiting example of a cytokine in this class that
causes the production of macrophages.   Thymic hormones both
natural and synthetic derivatives are another non-limiting
example of receptor ligand class of immune modulators, they are
an example of a subclass of receptor ligands that cause division
15   of immature immune mediator cells, in this case thymocytes or
premature lymphocytes.

Cytokines modulate target cells by interacting with cytokine
receptors on the target cell.   Principal cell sources of
cytokines include T lymphocytes, B lymphocytes, macrophages,
20   stromal cells, monocytes, leukocytes, and platelets.   While
cytokine specific receptors are specific for a given cytokine,
cytokine receptors are grouped into families based on shared
features.   The first group of cytokine receptors is the
hemopoetin group which include immune system cells that bind IL-
25   2, IL-3, IL-4, IL-6 and IL-7.   A second receptor family is the
TNF receptor family which bind both TNFα and TNFβ.   A third
family is the immunoglobulin (Ig) superfamily receptor family,
which contains an Ig sequence like motif and includes human IL-1
and IL-6 receptors.

See, e.g., Dawson, In *Lymphokines and Interleukins* (Dawson, ed.) CRC Press, Boca Raton, Florida (1991); Mosmann et al, *Immunol. Rev.* 123: 209-229 (1991); Mosmann et al, *Immunol. Today* 12:A59-A69 (1991); Sherry et al, *Curr. Opinion Immunol.* 3:56-60 (1991); Paul, *Blood* 77:1859-1870 (1991); Dower et al, *J. Clin. Immunol.* 10:289-299 (1990).

A second subclass in this receptor ligand group include lectins. A non-limiting example of a receptor ligand is a monoclonal antibody or fragment capable of binding and/or modulating a receptor, e.g., as a T cell receptor or an IL-2 receptor.

A second class of immune modulators are anti-receptor molecules. These agents can cause the production of antibodies or T cells that can either block receptors on a cell surface or kill such cells in a recipient. As non-limiting examples, one skilled in the art could induce active immunization in a recipient leading to the formation of antibodies or cytotoxic T cells that could, e.g., block lymphokine receptors on a cell, neutralize certain subtypes of antibodies, kill B lymphocytes that make certain subtypes of antibodies, or kill T lymphocytes that have a certain subtype of receptors on their surface.

A third class of immune modulators are "transplanted cells," which may include immune mediator cells as defined above that can induce responses by releasing lymphokines or by secreting other molecules. As non-limiting examples these cells can be lymphocytes, macrophages, splenocytes and/or thymocytes. As non-limiting examples the products the transplanted cells release can be lymphokines, as well as heterogenic or allogenic molecules as in proteins, carbohydrates and lipids.

A fourth class of immune modulators are "general immune modulators." These agents also go by other names including immune modulating agents or immune response modulators. Depending on what dose and how these agents are given they can be termed immune potentiators or immunosuppressants. These agents often have the ability to cause non-antigen specific activation of immune mediator cells through the release of lymphokines compared to vaccines that usually cause specific activation of clones with specific affinity for the vaccine antigens. General immune modulators are often used at higher doses than conventional immunogens and/or are typically given at frequent intervals as in every seven days or less. These agent often provide no protection against common pathogens in contrast to many vaccines and other pharmaceutically acceptable conventional immunogens. These agents generally do not employ adjuvants in contrast to vaccine immunogens. Non-limiting examples of immune modulators include the biologic OK-432 and the chemical entities levamisole and isoprinosine.

It should be noted that some agents may be members of more than one of the above groups or classes. As a non-limiting example, a lymphokine may be a conventional immunogen if it is derived from a heterologous species to which it is given as in the case of administering a mouse derived lymphokine to a human. Hybrid or fusion molecules can also be made that contain an biologically active part that has lymphokine activity and another section that contains an conventional immunogen.

IMMUNOGENIC AGENTS

An immunogenic agent (vaccine) of the present invention is a pharmaceutically-acceptable composition comprising at least one immunogen in an amount such that, when administered according to

5 an immunization schedule as disclosed above, it contributes to the desired effect against a chronic immune-mediated disorder, and, in a preferred embodiment, also against an infectious disease. In some embodiments, the immunogenic agent will comprise at least one non-pediatric immunogen and, optionally,

10 one or more pediatric immunogens. In a preferred embodiment, the immunogenic agent may comprise at least one anthrax immunogen, and at least one other immunogen, which may be a pediatric or another non-pediatric immunogen. In another preferred embodiment, the immunogenic agent may contain at least one plague

15 vaccine immunogen and at least one other immunogen, which may be a pediatric or another non-pediatric immunogen.

The term "immunogenic agent" is not intended to cover foods customarily given for nutritional reasons to infants and other children, such as bovine milk, common baby formula, and common

20 baby food, even though such foods may nominally contain "immunogens".

When multiple immunogens, and/or multiple dosings of the same immunogen, are administered, the individual doses of individual immunogens may by themselves be subimmunogenic,

25 provided that in aggregate, when administered according to the schedule, an immunogenic effect is achieved.

Immunogenic agents may be prepared in pharmaceutically
acceptable form according to known method steps, based on the
teaching and guidance presented herein. Non-limiting examples
of preparation of immunogenic agents suitable for use in the
present invention include DTP (21 CFR, part 610-630; Mueller et
al *J. Immunol.* 56:143 (1947)); tetanus/diphtheria Pillemer et
al., *J. Immunol.* 54:213 (1946)); measles/mumps/rubella (U.S. pat.
No. 4,147,772 to McAleer et al); yellow fever (*Morbidity and
Mortality Weekly Report* 27:268 (1978); *HHS Publication No. (CDC)
81-8280*, U.S. Gov't Printing Offc., Washington, D.C. (1981));
typhoid (*Morbidity and Mortality Weekly Report* 27:231 (1978));
plague (*Morbidity and Mortality Weekly Report* 27:231 (1978));
Hepatitis B (U.S. pat. No. 4,129,646, McAleer et al); hemophilus
((U.S. pat. No. 4,196,192, Kuo); OPV (Eagle, *Science* 122:501
(1955); anthrax (FDA Freedom of Information Act document
regarding Michigan Dept. of Health Anthrax Product License
Application, 1993), the entire contents of which are herein
entirely incorporated by reference.

Such immunogenic agents in pharmaceutically acceptable form
may also be prepared according to known method steps using
recombinant technology and by the use of monoclonal antibodies
and fragments thereof. See, e.g., Ausubel et al, eds, *Current
Protocols in Molecular Biology*, Greene Publishing Assoc. and
Wiley Interscience, N.Y., N.Y. (1987,1993); Coligan et al eds.,
Current Protocols in Immunology, Greene Publishing Assoc. and
Wiley Interscience, N.Y., N.Y. (1992, 1993); Sanbrook et al
infra, are Harlow, *infra*.

A hepatitis B vaccine may be recombinant or produced from
blood products. A polio immunogen may be a live and or killed

immunogen.   A polio vaccine may be trivalent such that the
vaccine may induce in mammals antibodies reactive to three
serotypes of polio virus.   A hemophilus influenza immunogen may
be a conjugated and or unconjugated immunogen.   A pertussis
immunogen may be a non-whole cell or a whole cell immunogen.   The
non-whole cell immunogen may include a cellular pertussis
immunogen.

Purification of vaccine immunogens helps minimize the
presence of impurities in a vaccine.   Purification may be
performed by any method that produces an highly pure end product
as in the non-limiting examples of: chromatography, electro-
kinetic processes, membrane processes, centrifugation, and
extraction (Biotechnology 5:789-793).   Extensive reviews of
separation techniques are included in several different texts
including the book by Juan A. Asenjo, Separation Process in
Biotechnology, published Marcel Dekker Inc., New York (1990) (see
especially Chapters 19, 20).   The preferred method of purifying
an non living vaccine antigen would be one that can inexpensively
produce highly pure end products with minimal loss of vaccine
immunogen in the process.   An example of a preferred method would
be purification using affinity chromatography where a column
contains a ligand that has high specificity, high binding
capacity for the target antigen and said antigen can be eluted
from said column under conditions that do not degrade the product
or the column.   In a preferred method a step should be added to
ensure the removal of fragments of the desired immunogen that
retain the ability to bind to the affinity column.

A vaccine should preferably contain as few impurities as
possible, as well as maintain a very high consistency of the

amounts and ratio of vaccine components including immunogens and adjuvants in each dose with little or no lot-to-lot variation. Vaccine immunogens should preferably be characterized by their unique physical properties to allow their composition in a

5 vaccine to be measured. The movement of an vaccine or its immunogenic components through a gel as in the non-limiting examples of chromatography or electrophoresis will allow detection of bands of impurities in the vaccine. In a preferred embodiment the total amount of impurities comprising molecules

10 unrelated to the desired vaccine components should be less than 0.5% by weight and even more preferably less than 0.1%. The desired vaccine antigens should preferably be defined by molecular weight and molecules should preferably differ by less than 5% of this number or be considered separate immunogen. The

15 variation in the amount of an particular defined vaccine compo-nent as in the non limiting examples of an immunogen or adjuvant in a dose should preferably be less than 2% by weight. A non-limiting example of how vaccines are currently tested for purity is described in a paper on manufacturing of Neisseria meningitis

20 vaccines (Avshalom Mizrahi, Bacterial Vaccines, Alan R. Liss Inc. (1990), pages 123-145) and differences to the current inventions are clear.

In addition to the immunogen, the pharmaceutical composition may contain suitable pharmaceutically acceptable carriers, such

25 as excipients, carriers and/or auxiliaries which facilitate processing of the active compounds into preparations which can be used pharmaceutically.

Such carriers may include depot adjuvants that release an immunogen *in vivo* over a prolonged period as compared to

administration of an unbound immunogen. Preferably the depot adjuvant comprises an aluminum, calcium or salts thereof, such as aluminum sulfate (alum), aluminum phosphate, calcium phosphate or aluminum hydroxide, see, e.g., Gregoriades, G. et al., *Immunological Adjuvants and Vaccines*, Plenum Press, New York, 1989. Depot adjuvants may prove especially useful in connection with diphtheria, tetanus, pertussis, hepatitis B, hemophilus influenza, and polio vaccines.

Another non-limiting example of a preferred ingredient is one that targets and/or activates macrophages, such as a liposome. See, Michalek, S.M. et al., Liposomes as Oral Adjuvants, *Curr. Top. Microbiol. Immunol.* 146:51-58 (1989).

Pharmaceutical compositions comprising at least one immunogen useful according to the present invention may also include suitable solutions for administration, intramuscularly, intravenously, subcutaneously, dermally, orally, mucosally, or rectally or by any other injection, and contain from about 0.001 to 99.999 percent, preferably from about 20 to 75 percent of active component (i.e. the immunogen) together with the excipient. Compositions which can be administered rectally include suppositories. Preparations of immunogenic agents for parenteral administration include sterile aqueous or non-aqueous solutions, suspensions, and emulsions, which may contain auxiliary agents or excipients, such as suitable adjuvants, which are known in the art. Pharmaceutical compositions such as tablets and capsules can also be prepared according to routine methods. See, e.g., Berker, *supra*, Goodman, *supra*, and Avery, *supra*, which are entirely incorporated herein by reference, included all references cited therein.

The immunogenic agents of the present invention may optionally include immunomodulators other than immunogens. Such immunomodulators may also be administered separately as a part of the program.

5   The compositions of the present invention may also include pharmaceuticals whose primary activity is non-immunological. Said non vaccine pharmaceutical agent may be of any class of pharmaceutical agents including the non limiting examples of agents to ameliorate the following diseases: infectious, cardio-

10   vascular, gastrointestinal, endocrine/hormonal, renal, neuro-logical, psychiatric, muscular, skeletal/orthopedic, hemato-logical, hepatic, pancreatic, metabolic, neoplastic, inflam-matory/rheumatic, reproductive, dietary/nutritional, ophthal-mologic, otologic, pulmonary/respiratory, dermatologic, allergic,

15   and surgical as in anesthetics. See Physicians Desk Reference, Medical Economics Data Production, Montvale, NJ (1994); United States Pharmacopeia; European Pharmacopoeia; Gilman et al, Goodman and Gilman's The Pharmacological Basis of Therapeutics, Macmillan Publishing Company, New York (1985). Preferably, these

20   pharmaceuticals are ones which, when screened as taught herein, are shown to help reduce the incidence or severity of a chronic immune-mediated disorder, or at least do not worsen the disorder.

The term "pharmaceutically acceptable" has both a legal definition and a clinical definition. The code of federal

25   regulations (21 CFR Part 600) legally defines what make an agent "pharmaceutically acceptable." Non-pharmaceutically acceptable products may be contaminated with agents, such as endotoxin or other pyrogens, the presence of which can have profound adverse effects on the immune system of the recipient. Results from

immunologic experiments using an non-pharmaceutically acceptable agent e.g., a lymphokine contaminated by endotoxin, may be quite different from an identical experiment using said lymphokine produced under GMP which lacks said impurities.

An immunogenic agent that meets the legal definition of "pharmaceutically acceptable" must further meet the clinical definition of "pharmaceutically acceptable". A clinical definition of pharmaceutically acceptable, as used herein, requires that said agent has a sufficiently beneficial clinical effect when used in a pharmaceutically acceptable dose.

A pharmaceutically acceptable dose, as in total dose, is a dose where the clinical benefits of said product outweighs the toxicity at said dose. Non-limiting examples of said toxicity include acute or subacute reactions like fever, shock or seizures, which may lead to permanent sequela and chronic toxicity like cancer, as is known and recognized in the relevant arts. A pharmaceutically acceptable dose according to this definition can vary according to the severity of the illness being modulated by the immunogenic agent. It is logical that a high dose of an agent which causes significant toxicity may be pharmaceutically acceptable for treating a life threatening malignancy yet the same dose would not be pharmaceutically acceptable for treating a benign disorder like a common sore throat. Following said logic, vaccination or other regimens to prevent diseases in healthy individuals usually employ non toxic doses. In the case of vaccinations, permanent sequelae as infrequent as one in 300,000 following immunization, as is believed the case with the whole cell pertussis vaccine, may be considered unsuitable by some.

A pharmaceutically acceptable dose will depend on the structure of the particular agent and/or the condition or genotype of the recipient. Some agents may be more toxic than others while some may be more immunogenic than others. In a like manner some individuals are more responsive to a given dose while others may be more sensitive to the toxic effects at that dose. There is thus an individual variation within the definition of pharmaceutically acceptable dose as well as species, racial, age, and population variation, all of which should be taken into account when dosing an individual. Such consideration has, of course, been given to other prophylactic agents.

The term pharmaceutically acceptable dose as defined herein may also incorporate an economic definition. A pharmaceutical acceptable dose in terms of preventing disease is one where benefit to society approximates or is greater than the cost of administering the agent. Said cost of administering the agent may include the cost of the agent, the necessary supplies as in needles, any transportation needed to bring patient, and the staff expenses needed to administer said agent, maintaining an administration clinic, as well as the costs associated with or resulting from adverse reactions to the agent. The benefits to society include savings from reductions in costs associated with the diseases that are prevented in society include one or more of lost productivity, medical expenses, expenses related to the care of the disabled person and some or all complications stemming from the illness. As a non-limiting example a pediatric vaccine that costs a society one trillion dollars a year to administer might not be considered pharmaceutically acceptable for a disease that costs said society ten million dollars a year.

A pharmaceutically acceptable dose must usually be determined by performing screening tests for both efficacy and safety. Data presented herein shows that doses of vaccines approximating that used to protect humans against infectious diseases (immunogenic doses) were also effective in preventing chronic immune mediated disorders in certain mammals. Pharmaceutically acceptable immunogenic doses for many of the preferred agents presented herein are thus readily available through common references with an nonlimiting example being the physician's desk reference, manufacturers product inserts, and scientific literature referenced through a database like MEDLINE. The immunogenic doses mentioned in the reference may be adjusted for the size of the recipient. A screening method as described herein may be used to calculate a pharmaceutically acceptable and immunogically acceptable dose.

The term pharmaceutically acceptable dose includes a supraimmunogenic dose. The lowest dose of immunogen effective in protecting the recipient against an infectious disease is termed the immunogenic dose. Using higher doses than the immunogenic dose results in a higher morbidity and mortality caused by the immunization than the additional morbidity and mortality, related to the infectious disease, prevented by using the higher dose. The current invention teaches that doses above the immunogenic dose may lead to enhanced protection against chronic immune mediated disorders.

A supraimmunogenic dose may be defined in several ways. One definition may be a dose that is higher than has been used to induce an protective immune response against the intended pathogen in recipients of a given age/size. The immunogenic dose

can be determined from art describing immunization against infectious diseases, as described herein. A second definition of supraimmunogenic dose may be a dose higher than necessary to prevent an infectious disease, where the enhanced toxicity associated with using the higher dose is greater than any measured benefit in preventing the intended infectious disease derived from using the higher dose. The toxicity and benefit may be determined by calculating the number of people expected to develop vaccine related toxicity and the number expected to be spared from the sequela of the infection disease that the vaccine prevents, where the toxicity and benefit may be quantitated into short term, moderate, and permanent sequelae. The toxicity and benefit may also be calculated financially as described herein.

It is understood that the dosage of an immunogenic agent of the present invention administered *in vivo* or *in vitro* will be dependent upon the age, sex, health, and weight of the recipient, kind of concurrent treatment, if any, frequency of treatment, and the nature of the effect desired. The ranges of effective doses provided below are not intended to limit the invention and represent preferred dose ranges. However, the most preferred dosage will be tailored to the individual subject, as is understood and determinable by one of skill in the art, without undue experimentation. In the context of the present invention "one dose" may include concurrent or separate administration of more than one immunogen comprised of an immunogenic agent according to the present invention. See, e.g., Berkow et al, eds., *The Merck Manual*, 16th edition, Merck and Co., Rahway, N.J., 1992; Goodman et al., eds., *Goodman and Gilman's The Pharmacological Basis of Therapeutics*, 8th edition, Pergamon

Press, Inc., Elmsford, N.Y., (1990); *Avery's Drug Treatment: Principles and Practice of Clinical Pharmacology and Therapeutics*, 3rd edition, ADIS Press, LTD., Williams and Wilkins, Baltimore, MD. (1987), which references and references cited therein, are entirely incorporated herein by reference.

The total dose, as in a pharmaceutically acceptable dose, required for each treatment may be administered by multiple doses or in a single dose. An immunogenic agent may be administered alone or in conjunction with other therapeutics directed to immunologic disorders, such as allergies, immune mediated cancers and autoimmune pathologies, as known in the art.

The pharmaceutically acceptable dosage of the immunogene will usually be about 0.01 μg to about 5 mg of immunogen, per kg body weight, and preferably from about 0.1 μg/kg to about 1 mg/kg body weight, still more preferably about 1 μg/kg to about 300 μg/kg, most preferably about 10 μg/kg to about 100 μg/kg. Nevertheless, Applicants' invention is not limited to the dosages set forth above. The active agent is the at least one immunogenic agent that induces an immune response according to the present invention. The safe dose will vary depending on the agent. Some immunogens are toxic at low doses while others are not.

KITS

The present invention also encompasses kits for administration of one or more immunogens according to methods of the present invention. Such kits may provide receptacles of immunogenic agents in forms suitable for pharmacologic administration, as described herein or as could be provided by

one skilled in the art based on the teaching and guidance presented herein, without undue experimentation. Preferred examples of such forms for administration are for oral or injection administration. Such kits may include, as non-limiting examples, at least one immunogenic agent comprising an anthrax or plague immunogen and at least one other immunogen corresponding to a different organism and or disease selected from the group consisting of diphtheria, pertussis, measles, mumps, rubella, small pox, pneumococcal, CMV, HIV, meningitis, tetanus, hemophilus influenza, hepatitis B, hepatitis, cholera, varicella, typhoid, yellow fever, neisseria, plague, adenovirus, hepatitis C, herpes, anthrax, plague, malaria, rabies, and/or polio virus immunogen, or a molecule that cross reacts immunologically to the immunogens. Another non-limiting example of such kits includes at least one non-pediatric immunogen and at least pediatric immunogen. Optionally and preferably the kits are in the form of a pharmaceutical composition optionally further comprising a pharmaceutically acceptable carrier, such as a vaccine adjuvant or a depot adjuvant, such as aluminum, calcium or a salt thereof.

PHARMACEUTICAL ADMINISTRATION

The immunogenic agents of the present invention may be administered by any effective route, for example, by various parenteral routes such as subcutaneous, intravenous, intradermal, intramuscular, intraperitoneal, intranasal, transdermal, or buccal routes. Alternatively, or concurrently, administration may be by the oral route. A preferred mode of using an immunogenic agent or composition of the present invention is by intramuscular application.

SCREENING METHODS FOR DETERMINATION OF WHETHER PARTICULAR IMMUNOGENS, IMMUNOGENIC AGENTS AND/OR IMMUNIZATION SCHEDULES AFFECT THE INCIDENCE OF SEVERITY OF A CHRONIC IMMUNE-MEDIATED DISORDER.

5   The present invention also provides methods for screening potentially pharmaceutically acceptable immunogenic agents and or dosing regimens for their suitability and effectiveness in preventing at least one chronic immune mediated disorder and at least one infectious disease. One skilled in the art can perform 10   any of the following screening methods or as few as desired to determine information sought by the investigator. The preferred order of performing the screening methods is listed below, however, one can perform the methods in any order that best suits the investigator. The term potentially pharma-ceutically 15   acceptable implies that the agents are not associated with any toxicities, at doses that they are being used in the screening methods, that would prohibit their use.

The methods may be adapted to screen for the ability of a immunization schedule containing one or more potentially 20   pharmaceutically acceptable immunogens and, optionally, one or more potentially pharmaceutically acceptable immune modulators, to prevent both chronic immune mediated diseases and infectious diseases. Immune modulators may be substituted for one or more immunogens in the following methods where applicable. Appropriate 25   doses of immune modulators to use can be calculated from references cited herein, references found in Medline/Index Medicus or other suitable database, and for new immune modulators one can use doses by weight approximating those used for immunogens.

Suitable screening methods, such as described herein, will allow one skilled in the art to determine (a) if a substance is an immunogen; (b) if it is effective in preventing disease; (c) if it has a safe and effective range; and (d) if said agent

5   modulates at least one chronic immune mediated disorder in mammals that already receive immunogens early in life to protect against infectious diseases. For immunogens thus shown to be useful, one may further determine a safe and effective immunogenic agent and/or dosing regimen utilizing said immunogen,

10   alone or with others. After an effective immunization schedule is determined, the screening methods may be used to verify the effectiveness of vaccine lots for the purpose of preventing or alleviating a chronic immune-mediated disorder.

Alternatively, a screening trial may be designed to

15   determine if an immunization schedule, such as a standard schedule known in the art, will induce and/or enhance the incidence and/or severity of at least one chronic immune mediated disorder. In the latter case, it may be especially useful in screening production lots of approved vaccines for the hitherto

20   unrecognized safety problem of inducing or exacerbating a chronic immune-mediated disorder.

The preferred method for finding immunogens suitable for use in the present invention is to start with an extract derived from at least one foreign organism, such as a bacteria, virus, fungus,

25   insect, plant, allogenic or xenogenic animal. A simple screening test for determining if an agent is an immunogen is to inject the agent subcutaneously in an mammal, such as a mouse, and looking for the development of inflammation at the injection site as well as the development of antibodies to the agent. Suitable sites

include paw pads and ears. Mammals that appear to have an immune response can have blood drawn and checked for anti-bodies to the agent. An ELISA is an simple assay that can be used to screen many animals for antibodies very quickly. Delayed type

5   hypersensitivity assay can be performed for determining if the agent induces a cellular immune response. Controls can be made by using animals that receive only the vehicle used to suspend the potential immunogen in question.

In vitro assays can also be developed to screen for poten-

10   tial immunogens. These assays are best suited for screening immune modulators like lymphokines, and certain monoclonal antibodies believed to have immune regulatory effects. As a non-limiting example, splenocytes, thymocytes or lymphocytes can be cultured in a 96 well plate with supportive media. Uptake of

15   tritium labeled thymidine 48-72 hours after the addition of an potential immunogen can suggest the activation and proliferation of immune mediator cells, suggesting that said agent is an immunogen.

Once an agent has been confirmed to be an immunogen, a

20   screening study can be performed to determine its effectiveness in preventing at least one chronic immune mediated disorder. High doses of the immunogen are recommended initially so that an effect due to using too low a dose is not missed. Preferred starting dose is 1 to 2 mg/kg body weight. A preferred method

25   is to give autoimmune prone mammals intraperitoneal injections with the agent starting within 10 days of birth and continuing every 2 weeks for 4 injections. Control animals can be injected with phosphate buffered saline or any vehicle that the immunogen is dissolved in. Blood can be drawn from the mammals and checked

for the presence of hyperglycemia in diabetic prone mammals or autoantibodies in autoimmune prone mammals that develop antibody mediated disease. Tissue from the study mammals can be removed and histology can be performed to determine if the tissue is

5 affected by chronic immune mediated disorders. Non-limiting examples include pancreatic islet cells in diabetes prone mammals or gastric fundi in mammals prone to develop autoimmune gastritis. Histology is generally a less sensitive assay than ELISA assays for autoantibodies or blood glucose for diabetes.

10 All results in the treated group can be compared to the control mammals.

Preferably, immunogens are screened for the ability to react to the recipient's sera. Preferably the sera from individuals, not pooled, should be screened. Many methods have been developed

15 to determine if antibodies react to an antigen. Any method that performs the desired function is suitable. Antibody titers can be determined to quantitate the antibodies in sera that react to the immunogen. As an alternative method, maternal serum, from the potential recipient's mother, may be screened since the

20 recipient often receives maternal antibodies which cross the placenta.

Several screening methods lend themselves very well to determining if the agent has effects on inhibiting chronic immune mediated disorders. Non-obese diabetic (NOD) mice or Bio

25 Breeding (BB) rats can be used to screen an agent for effectiveness in preventing diabetes. Mice can be injected as above and blood drawn to check for hyperglycemia at monthly intervals through the first 6 months of life. Ten or more female NOD mice will give an quick first approximation of an effect.

Detailed methods are provided in examples 1 and 2 below. Other possible methods of screening agents for their effectiveness in preventing chronic immune mediated diseases beside diabetes also exist. Cyclosporine induced autoimmunity, e.g., as demonstrated
5   in example 3, is one method for screening agents for their ability to inhibit organ specific autoimmune disease like gastritis. Different strains of mice are more prone to develop certain autoimmune diseases when they are altered then others (Kojima and Prehm, 1981). One can pick a strain depending on
10   what disease one wants to look at.

Other immunosuppressing regimens besides cyclosporine can be used to create disease in animals that can be inhibited by vaccination. Non-limiting examples include immunosuppressive monoclonal antibodies (Journal of Experimental Medicine 166:461-
15   475), other noncyclosporine immunosuppressants should preferably be given for the first week of life as in Example 3 and then vaccines may be administered. Other rodents that can be used to screen for vaccine prevention of autoimmunity include the non-limiting examples of MRL/1pr (see Example 5) and NZB/NZW mice
20   which spontaneously develop rheumatoid diseases that resemble lupus and rheumatoid arthritis.

Blood can be screened for autoantibodies using ELISA assays as described in example 3 below. There is a large number of ELISA assays that have been published in the scientific
25   literature and these can be found looking through index medicus or a similar computer file referencing autoantibodies and the particular autoantigen or organ system one wants to study. See also Ausubel, *supra*, Coligan, *supra,* Harlow, *supra*.

One skilled in the art may determine if an agent can alter the incidence, prevalence, frequency and or severity of a chronic immune mediated disorder by studying the effect of said agent on surrogate markers of said disorder. Theses markers may include

5   the presence of lymphokines, autoantibodies, markers of cell destruction as in the non limiting example of increased levels of certain enzymes in the blood, indicators of abnormal endocrine function as in the non limiting example of abnormal glucose tolerance test or thyroid function test, autoimmune lymphocyte

10 clones, certain immune mediator or inflammatory cells associated with increased risk of autoimmunity which correlate with toxicity. See Isselbacher et al. editors, Harrison's Principles of Internal Medicine, McGraw-Hill, New York (1994); Rose, N.R et al. editors, Manual of Clinical Laboratory Immunology, American

15 Society for Microbiology, Washington, D.C. (1992); Roitt, Ivan Maurice et al. editor, Encyclopedia of Immunology, Academic Press, London (1992); Miller, ·Linda E. et al., Manual of Laboratory Immunology, Lea & Febiger, Philadelphia (1991); Bona, Constantin A. et al.; Molecular Immunobiology of Self-reactivity,

20 Dekker, New York (1992), Talal, Norman, Molecular Autoimmunity, Academic Press London (1991); Bouloux, Pierre-M. G. et al., Diagnostic Tests in Endocrinology and Diabetes, Chapman & Hall Medicine, London (1994); Noe, Dennis A et al., Laboratory Medicine: The Selection and Interpretation of Clinical Laboratory

25 Studies, Williams & Wilkins, Baltimore (1994); Tilton, Richard C. et al., Clinical Laboratory Medicine,. Mosby Year Book, St. Louis , (1992); Paul William E., Fundamental Immunology, Raven Press, New York (1993).

Agents that have been shown to effectively inhibit chronic immune mediated disorders at high doses may be screened to determine what doses if any may be safely administered to an mammal. Routine toxicology tests, as those requested by a regulatory agency such as the FDA, can be performed at varying doses to determine an safe dosing range. Non limiting examples of such tests include mutagenesis, carcinogenesis, acute toxicity on vital organs such as the kidneys, lung, heart, liver, and brain, chronic toxicity of the organs, as well as defining a lethal dose.

The present invention also pertains to a method to screen vaccines to determine their propensity to induce diabetes mellitus or chronic immune mediated disorders like autoimmune diseases. A group of autoimmune prone mammals may be given an agent in the first months of life, as an non-limiting example, a less strong immunogen, to reduce the incidence of autoimmunity in those mammals. A diphtheria/tetanus vaccine is a preferred embodiment and the dose used is expected to reduce the incidence of autoimmunity by 50%-75% compared to untreated mammals, but should preferably not completely inhibit autoimmunity. A second group of autoimmune prone mammals should receive the same immunogen and dose, but should receive the vaccine in question at approximately 2 months of age or later, with 2 months preferred. The incidence of diabetes or autoimmunity should be noted in each group. Safety should be assessed by determining the incidence or prevalence of disease in the two groups. An example of the method is demonstrated in example 2, where the diphtheria tetanus vaccine is given to 2 groups of NOD mice but one group also received an additional administration of pertussis

vaccine at approximately 2 months of age. The later group developed a higher incidence of diabetes suggesting that pertussis vaccine can have a toxic effect by inducing diabetes. Non limiting examples of other possible autoimmune prone mammals

5   that can be used in the screening method include cyclosporine-treated mice, see Example 3 below, and day-3 thymectomized mice.

The mammals in the previous methods should be raised in an pathogen-free environment as a preferred embodiment. A non limiting example of an suitable environment includes housing

10   animals where the caging, bedding, food and water are all autoclaved, the cage has filters to prevent pathogens from entering the cage. As an preferred environment the cage should only be opened under a laminar flow hood equipped with filters to trap pathogens. The use of an pathogen free environment

15   prevents mammals from exposure to natural immunogens that may effect the interpretation of the results.

The present invention also pertains to a method to screen potentially pharmaceutically acceptable immunogens to determine their propensity to modulate at least one chronic immune mediated

20   disease in mammals that live in open environments, as opposed to pathogen free environments, which receive extensive exposure to both pharmaceutically acceptable and natural immunogens early in life. In said method, mammals will typically receive at least one vaccine at less than 96 months of age where the agent is

25   capable of preventing an infectious disease caused by an infectious agent or capable of inducing neutralizing antibodies to the infectious agent. In the screening method the dosing regiment to prevent at least one chronic immune mediated disorder

and at least one infectious agent is termed an immunization protocol.

The screening method may involve administering at least one potentially pharmaceutically acceptable dose of at least two potentially pharmaceutically acceptable immunogenic agents comprising at least one potentially pharmaceutically acceptable pediatric immunogen and at least one agent selected from the group consisting of a second pediatric immunogen and a non-pediatric immunogen, for the ability to modulate the development of at least one chronic immune mediated disorder and or of at least one surrogate marker of the chronic immune mediated disorder in a population and or subpopulation of mammals. The method comprising administrating to at least one treatment group of at least one mammal at risk for at least one chronic immune mediated disorder at least one treatment dose of immunogenic agents according to a treatment administration schedule. The at least one treatment dose may comprise at least a first treatment dose of at least one of the immunogenic agents administered prior to an age of 56 days of the mammal.

One or more control groups of at least one mammal may be defined optionally. The control groups may receive at least one control dose which contains at least one potentially pharma-ceutically acceptable pediatric vaccine and has at least one modification selected from the group consisting of: (i) lacking at least one immunogenic agent/adjuvant than as is provided in the treatment schedule; (ii) including at least one immunogenic agent/adjuvant than as is provided in the treatment schedule; (iii) including a higher dose of at least one immunogenic agent/adjuvant than as is provided in the treatment schedule;

(iv) including a lower dose of at least one immunogenic agent/adjuvant than as is provided in the treatment schedule; (v) including at least one additional dose of at least one immunogenic agent/adjuvant than as is provided in the treatment schedule; (vi) lacking at least one dose of at least one immunogenic agent/adjuvant than as is provided in the treatment schedule; (vii) including at least one dose of at least one immunogenic agent/adjuvant at a later time than the immunogenic agent/adjuvant is administered in the treatment schedule; (viii) including at least one dose of at least one immunogenic agent/adjuvant at an earlier time than the immunogenic agent/adjuvant is administered in the treatment schedule; and (ix) no modifications from the treatment schedule wherein at least one measure in the group consisting of incidence, prevalence, frequency, and severity of at least one chronic immune mediated disorder and or of at least one surrogate marker of the disorder can be/has been determined in the control group. The term immunogenic agent in this paragraph refers to an immunogen that is unbound or administered with a particular pharmaceutical carrier at a definable ratio as described herein. The terms treatment schedule and treatment administration schedule are used interchangeably herein.

Participants may be screened to determine the modulation of development of the at least one chronic immune mediated disorder by at least one of the treatment administration schedules. The determining step may comprise ascertaining at least one measure from the group consisting of incidence, prevalence, frequency and severity of at least one chronic immune mediated disorder and or of at least one surrogate marker of the disorder in at least one

mammal in at least one treatment group and optionally the same measures in at least one mammal in the one or more control groups. Historical controls may be used in place of experimental controls.

5   A control group in one screening trial may be the treatment group in the next or vice versa. In a preferred embodiment, a control group is used which contains mammals that receive an currently recommended immunization schedule as in the non-limiting examples of the schedules recommended by the Centers for

10   Disease Control (CDC) and the American Academy of Pediatrics.

Comparison of data from the one or more control groups to data from the at least one treatment group may be performed to determine if the treatment schedule can modulate at least one measure in at least one chronic immune mediated disorder. In a

15   preferred embodiment the size of the groups should allow determination of statistical significance during an acceptable period of time. Any statistical method that is deemed appropriate for the trial design by one skilled in the art may be acceptable.

In order to select an appropriate size of the treatment and

20   control groups it is first helpful to know the incidence/prevalence/frequency/severity of the chronic immune mediated disorder and or surrogate marker of the disorder in children. Given the desired change in incidence of disease one hopes to be able to detect, and the statistical test one is employing, one can

25   determine the size of a population needed for the at least one treatment group and the one or more control groups to demonstrate statistical significance ($p <$ or $= 0.05$).

It is preferred to perform the screening method in a developed country where the exposure to pathogens is reduced

during the first year of life. Developed countries have expensive water purification and sewer systems that reduce an infant's exposure to the pathogens. The method should preferably allow one skilled in the art to determine if the treatment administration schedule prevents, induces, or has no affect over the currently recommended immunization schedule.

Developed countries may include the United States, Canada, and the European nations listed in the table pertaining to Example 101. A country may be considered developed if it has both a per capita income greater than or equal to any of the above countries, and nationwide water purification systems as well as sewage treatment systems which equal or surpass that in the above nations.

The mammals of the screening methods may include as non limiting examples rodents such as NOD mice and BB rats, and primates such as humans.

A human study population will preferably contain individuals who are at risk for development of the chronic immune mediated disorder. For example, in the case of diabetes mellitus, it will preferably include at least 25% Caucasians of European descent. Caucasians of European descent have a high rate of diabetes and certain autoimmune diseases as compared to other groups like Japanese and this invention shows that occurrence of diabetes in European caucasians is sensitive to vaccination.

The screening method may be retrospective, prospective or contain data derived both retrospectively and prospectively. Individual mammals may be placed in a particular study group based a selection process that is either random, non random or both, however random selection is the preferred method.

Participants, their families, and researchers may be either blinded, unblinded or both to the immunization schedule. The endpoints of the screening method may be the diagnosis of at least one chronic immune mediated disorder and or surrogate marker of the disease in an individual. The preferred embodiment is to use at least one disease that develops early in life, is relatively frequent, and has a clearly defined endpoint with the non limiting example being type I diabetes mellitus.

By using the combination of several diseases as an end point the number of participants needed to show a statistical effect is reduced and the time to detect an effect is reduced. Prior to this invention it was not accepted that pharmaceutical acceptable doses of pharmaceutically acceptable immunogenic agents could prevent one chronic immune mediated disorder, much less several such disorders.

It is often easier to compare people in different countries because most people in a given country tend to receive the same vaccines, so it is hard to get an unbiased division of those who received from those that did not receive vaccine. Governments' decisions on whether to promote immunization of the citizens with a particular vaccine like BCG is often ambiguous. Thus one can have a situation where genetically and culturally similar people live on opposite sides of a national border and one group of citizens receives a particular vaccine and another nationality does not receive the vaccine or receives it at a different age.

Another non-limiting example of using retrospective data to screen an pharmaceutically acceptable immunogen for the ability to modulate a chronic immune mediated disease is to study the effect of changing an immunization schedule on the incidence of

disease. In places where the immunization schedule has changed as in the case of Finland, see Example 101 discussed herein, one skilled in the art can study the effect of the change on the development of at least one chronic immune mediated disorders.

5 The incidence/prevalence/frequency/ severity of the at least one disorder and or surrogate marker of the at least one disorder prior to the change may be determined retrospectively and or prospectively and the parameters of the at least one disorder after the change may be determined prospectively and or

10 retrospectively.

One skilled in the art may also perform prospective screening methods. As a non-limiting example mammals in a treatment group may receive one immunization schedule and mammals in a control group may receive a different immunization schedule

15 as described above. The incidence/prevalence/ frequency/severity of at least one chronic immune mediated disorder and or at least one surrogate marker of the at least one disorder is determined prospectively in each group. A nonlimiting alternative to the example is where the data from one group may be determined

20 retrospectively and the data from another is determined prospectively.

The screening method may be designed to control for and or estimate the effects on at least one mammal in at least one treatment/control group from at least one confounding variable.

25 For example, one may consider the effects of receiving breast feeding versus bottle feeding prior to 12 months of age, receiving antibiotics during the first 12 months of life, the maternal age at birth, the presence of a chronic immune mediated disorder in the mother/father/a close relative, maternal

infections while the mammal was in utero, infections during the first 12 months of life, size of the mammal at birth, gestational age of the mammal at birth, exposure of the mammal to BCG vaccine or naturally acquired mycobacterium, and exposure to various

5   vaccines or pathogens.

Studies may give misleading results on a vaccine's safety if one does not take into consideration that having a disease like measles may be associated with a higher incidence of a chronic immune mediated disorder than receiving the vaccine for

10   the disease. A vaccine may induce chronic immune mediated disorders but at a lower rate than the infectious agent it protects against.

One may design the trial to control for and or determine the difference in at least one chronic immune mediated disorder

15   between a group which received a vaccine but did not develop the infectious disease which the vaccine protects and a group that did not receive the vaccine but also did not develop the infectious disease. The difference in at least one chronic immune mediated disorder may include at least one measure in the group

20   consisting of incidence, prevalence, frequency, and severity of at least one chronic immune mediated disorder and or of at least one surrogate marker of the at least one disorder. Type I diabetes mellitus is the preferred chronic immune mediated disease to study for reasons mentioned above.

25   The optional one or more control groups may receive an administration of at least one potentially pharmaceutically acceptable pediatric vaccine prior to the age of 56 days of the at least one mammal.

The treatment group may receive at least one dose of at least one immunogenic agent/adjuvant not administered to at least one control group.

One may use several different methods to determine if an
5   mammal develops protection against at least one infectious disease for which the at least one mammal received immunization against in the pediatric age as defined earlier. In a non-limiting example one can determine the incidence/prevalence/frequency/severity of the at least one infectious disease in the at
10   least one treatment/control group in question and determine if the group received adequate protection from the at least one infectious disease. In an alternative method members of the group in question may be screened for the development of a protective level of antibodies to the causative infectious
15   agents. As a non limiting example the level of protection may be compared to at least one of the following: (i) a group that received no protection/immunization, (ii) a group that receives a recommended immunization schedule, (iii) an rate that is deemed acceptable by professionals in the field as in those affiliated
20   with the Centers for Disease Control, the American Academy of Pediatrics and or the FDA.

The size of a group needed to determine if an dosing schedule provides protection against at least one infectious agent is generally smaller than that needed to detect an effect
25   on reducing a chronic immune mediated disorder because the incidence of the former diseases are generally higher than the later diseases and the former may occur earlier in life than the later. If the end point of the study is the development of protective or neutralizing antibodies two hundred mammals or less

may be sufficient. The trial preferably will utilize any statistical method that is acceptable and is congruent with the study design.

Methods of the present invention may also be used to screen
5 non-vaccine pharmaceutical agents for either their ability to induce a chronic immune-mediated disorder including spontaneous immune-mediated diabetes mellitus and/or to interfere with a vaccines' effect on a chronic immune-mediated disorder. One skilled in the art will be readily able to adapt these methods
10 to perform the desired task, by substituting the non-vaccine pharmaceutical for the vaccine in the above-methods.

However, one need not determine if said pharmaceutical prevents at least one infectious disease. In the case where said pharmaceutical is screened for the ability to interfere with a
15 vaccines' effect on a chronic immune-mediated disorder, said pharmaceutical may be given with a vaccine and the results can be compared to an otherwise identical experiment where only the vaccine is administered. In a prefered embodiment, the chronic immune-mediated disease is diabetes mellitus. Said non-vaccine
20 pharmaceutical agent may be of any class of pharmaecutical agents. The pharmaceutical product may be one other than an immunosuppressant, an agent to regulate blood sugar, and/or an immune stimulant.

Methods and products for late and booster immunization

25   The present invention also encompasses compositions and methods to administer vaccines after 8 weeks of age without significantly inducing an chronic immune-mediated disorder. Animal and human epidemiology data described herein show that the

administration of vaccines starting after 8 weeks of life can induce chronic immune-mediated disorders including type I diabetes mellitus. Currently there are no ways of starting immunization after 8 weeks or providing boosters to older

5   children without a substantial risk of inducing diabetes. Compositions and methods described below may preferably be used for starting immunization after 8 weeks of age, administering a booster when the mammal has not received a vaccine in the prior 3 weeks, and administering vaccines when the recommended

10   administration schedule in the first 4 months of life was not complete.

Preferably, the subject is first screened for the adequacy of his or her protection against the infectious diseases against which immunization was considered desirable; if adequate,

15   immunization is contraindicated as it could increase the risk of inducing or exacerbating a chronic immune-mediated disorder.

Next, the subject is preferably screened for the presence of a clinical or sub-clinical chronic immune-mediated disorder, positive subjects are not good candiates for immunization that

20   could worsen their condition.

If the subject is still to be first immunized with a particular immunogen after eight weeks of age, the risk of thereby eliciting a chronic immune-mediated disorder can be blunted by using weaker immunogenic agents, administering an

25   immunosuppressant, or administrating a glucocorticoid hormone (or analogue), or a substance which induces glucocorticoid release.

Screening for a protective immune response and a chronic immune mediated diseases may be performed in sequence or jointly. Body fluids may be screened for the presence of antibodies in

levels indicating said mammal has sufficient protection against an infectious disease and if the mammal does not have sufficient protection then said body fluids may be tested for indicators of at least one chronic immune mediated disorder and or risks of
5 developing said disorder. Alternatively body fluids can be screened for both the presence of antibodies in levels indicating said mammal has sufficient protection against said infectious disease and for indicators of at least one chronic immune mediated disorder and or risks of developing said disorder. The
10 preferred body fluid is blood or a derivative of blood as in serum or plasma. Other body fluids that may be used include as non limiting examples saliva, urine, tears, and lymph. Said indicators of at least one chronic immune mediated disease and or risks of developing said disorder may include the following
15 non limiting examples: autoantibodies, lymphocytes containing certain T cell receptors, autoimmune lymphocyte clones, certain MHC types associated with increased risks for developing autoimmunity, certain immune mediator or inflammatory cells associated with increased risk of autoimmunity, an abnormal
20 glucose tolerance, ongoing tissue destruction, and endocrine abnormalities.

Screening may be performed in a clinic and or physician's office to give immediate results or an specimen as in the preferred embodiment but non limiting example of blood may be
25 removed from the patient for analysis at a later time or different sight. Screening may measure humoral immune responses or cellular immune responses. The latex bead agglutination test is a preferred method of detecting humoral immunity and auto-antibodies because it is fast, sensitive, specific, simple, and

inexpensive. Other non limiting examples of acceptable ways to detect antibody levels include ouchterlony analysis, radial immunodiffusion, electroimmunodiffusion, immunoelectrophoresis, radioimmunoassays, ELISA, complement fixation tests, other

5   agglutination reactions, cytotoxic assays, immunfloresecence, radioactive binding assays, immunoferritin or other electron-microscopy techniques, flow cytometry, hemolytic plaque assays. The preferred method of testing cellular immunity is through skin testing however more complicated techniques using cell culture

10   would also work. In a preferred embodiment the test are performed from a single sample of blood however several blood specimens taken from the patient in a short time span as in less than 72 hour period would also be readily acceptable.

        The methods above can be utilized by one skilled in the art

15   to determine the degree of protection against an infectious disease and or the risk of developing a chronic immune mediated disease. One skilled in the art as in the non limiting example of a physician may use this data to determine appropriate administration of vaccines. The following are non limiting

20   examples of how these methods may be applied. Antibody titers may provide information on the degree of protection against infectious diseases and the presence of underlying autoimmunity. An individual with no autoantibodies and high titers of antibodies protective against an infectious disease may receive

25   no immunization, an similar individual with moderate titers providing protection against an infectious disease may only receive one dose of vaccine, while an individual with no antibodies to the organism or its toxin may receive several doses of vaccine. An individual with an abnormal glucose tolerance test

and or autoantibodies against islet cells may not receive any vaccine irrespective of his or her risk for developing the infectious disease. An individual who develops high titers of autoantibodies after receiving a single dose of vaccine may be prohibited from receiving more vaccines.

The present invention also encompasses kits and products to perform the above mentioned screening methods. The kits and products may utilize any screening method suitable to perform the desired function. The following are several nonlimiting examples of products that could employ the screening methods. A product to determine if an mammal should receive an immunization where the product performs at least one of following: determining if said mammal has a protective immune response to a vaccine preventible organism and determining if said individual is at high risk for developing a chronic immune mediated disease that could be induced and or exacerbated by vaccination. A kit containing said product and a vaccine.

Methods and products described above may be beneficial for testing mammals in most age groups. The method is preferably employed prior to receiving each administration of vaccine when said mammal is older than 2 months of age especially if said mammal has not received a vaccine in the prior 3 weeks. The method may be used prior to receiving any vaccine immunogen as in the non limiting examples of an polio, BCG, smallpox, hepatitis B, diphtheria/tetanus/pertussis, an conjugated or unconjugated Hemophilus influenza, anthrax, plague, encephalitis, meningococcal, meningitis, pneumococcus, pneumonia, typhus, typhoid fever, streptococcal, staphylococcus, neisseria, lyme disease, cholera, E. coli, shigella, leishmania, leprosy,

cytomegalovirus, respiratory syncytial virus, Epstein Barr virus, herpes, influenza, parainfluenza, rotavirus, adenovirus, HIV, hepatitis A , NonA NonB hepatitis, varicella, rabies, yellow fever, measles, mumps, rubella, HTLV, rabies, Japanese
5   encephalitis, flavivirus, dengue toxoplasmosis, coccidiomycosis, schistosomiasis, malaria immunogen, and a molecule that cross reacts immunologically to at least one of said immunogen.

The diseases to which protection should be measured will depend on what vaccines are approved for marketing. The disease may be
10   one other than tuberculosis or rubella. As new vaccines enter the market then protective antibodies to their corresponding diseases may be measured. The vaccine may be a non living vaccine.

A screening method may be performed not only to determine if the antibody titers are protective but also to determine what
15   brand of vaccine the recipient received if any by looking for antibodies to particular brand specific antigens as in the nonlimiting example of detecting antibodies to a carrier protein incorporated in a Hemophilus influenza vaccine. It is important that an individual who received an initial series of immunization
20   with vaccines from one manufacture receives booster vaccines manufactured from that same manufacturer, preferably the same lot in order to prevent exposure to new antigens after 8 weeks. For example some Hemophilus influenza vaccines typically contain bacterial carbohydrate however some are not conjugated while
25   others are conjugated. One brand may be conjugated with a tetanus toxoid another with a different protein. A vaccine registry would be ideal however people's name can not always be found on these registries. A test kit to determine the brand of vaccine the recipient received would help reduce the incidence of diabetes.

A vaccine manufacturer may also include manufacturer specific immunological markers in their vaccines to help allow differentiation of brands of vaccines.

The present invention also encompasses methods, vaccines and vaccine kits that have a decreased propensity to induce a chronic immune mediated disorder when administered to an mammal of at least 2 months of age. These vaccines and vaccine kits may comprise vaccines and a immunosuppressant. The risk of inducing a chronic immune mediated disorder during booster administration may be lowered by boosting with an vaccine which lacks a strong adjuvant and using small molecular weight immunogens which retain the protective epitopes but induce less inflammation and are less immunogenic. An adjuvant like an aluminum salt may activate macrophages causing increased risk of type I diabetes mellitus and other chronic immune mediated diseases. Using a non living vaccine that lacks an aluminum based adjuvant may decrease this effect. Administration of an immunosuppressant preferably a glucocorticosteroid like cortisol or a longer acting version will help decrease inflammation including macrophage activation and decrease the risk as well. A hormone or other molecule capable of inducing the release of glucocorticoids as in the non limiting example of adrenocorticotropic hormone (ACTH) and corticotropin releasing hormone (CRH) would also perform the same function. A list of glucocorticoids and other immunosuppressants as well as their recommended administration are described in the Physician's Desk Reference 1994. Administration doses will vary from recipient by recipient as known by one skilled in the art and administration may be optimized without unnecessary experimentation by using dosing instruction from readily available sources

including the Physician's Desk Reference and information supplied herein especially the section on screening methods.

Having now generally described the invention, the same will be more readily understood through reference to the following examples which are provided by way of illustration, and not intended to be limiting of the present invention.

EXAMPLES

In Examples 1 and 2 mentioned below, initial vaccinations were either at day 8, 15 or day 1, 3, 10 respectively. This initial schedule was designed to minimize the phenomenon of under immunizing mice due to extravasation of vaccine from their abdomen and inadvertent injection into the visceral organs, bladder and gastrointestinal tract, which results in rapid clearance of the vaccine. Less frequent early administrations are needed for larger mammals when one is sure that extravasation or visceral injection does not occur. After the initial challenge injections every other week or even less frequent suffice.

The method of administration and the compositions contained in this invention allow relatively low amounts of immunogenic agent to be given. The anthrax vaccine, for example, as supplied by the manufacturer contains less than 200 micrograms of protein/ml as calculated using Lawry-based protein determination kit (Sigma, St. Louis, MO). Using the dilutions given in Example 1, the amount injected into mice comes to 0.2, 0.6, 0.8 μg protein on day 8, 15, 29. These amounts correspond to roughly 40-60 μg/kg body weight which are pharmaceutically acceptable doses.

The DT vaccine used contains about 180 micrograms of protein per ml and the DTP vaccine contains about 544 micrograms per ml.

The following examples involve immune modulation using commonly available human vaccines: anthrax vaccine, A, (Michigan Department of Health, Lansing, Michigan); combined diphtheria, tetanus vaccine, DT, (Connaught, Swiftwater, PA); combined whole cell diphtheria tetanus pertussis vaccine, DTP, (Connaught); and plague vaccine, P, (Miles, West Haven, CT). All vaccines are produced from killed microorganisms, and all except the plague vaccine have an aluminum based adjuvant. The DT vaccine contains purified toxoids while the anthrax and pertussis vaccines are less pure. Vaccines are diluted using sterile technique in sterile phosphate buffer solution pH 7.4 using the notation 1:100 to indicate 1 part vaccine per 100 part PBS by volume. Vaccines were administered by intraperitoneal injections for convenience though other routes would be effective.

It is known that even though vaccines from different manufacturers provided similar protection to an infectious agent different vaccines contain different antigens, and different amounts of similar antigens (Merk Sharp Dome 1991). The following data shows that the exposure to new antigens at 2 months increases the risk of developing diabetes and chronic immune mediated disorders. The epidemiology data shown below demonstrates that different vaccines can interact to decrease the incidence of chronic immune mediated disorders or increase them.

Examples 1, 2 and 4 describe the anti-diabetic effect of the immunization programs herein disclosed on diabetes in NOD mice (exs. 1 and 2) and diabetes in BB rats (ex. 4). It is now widely accepted by those skilled in the art that type I diabetes in

humans responds similarly to immune intervention as does diabetes in NOD mice and BB rats. Diabetes in all three species is considered to be an autoimmune disease based on the presence of islet cell autoantibodies and strong genetic linkage between the

5 development of diabetes and MHC genes (New England Journal of Medicine 314:1360-1368,1986; Diabetes Reviews 1:15-42,1993). Immunological events occurring in the first 2 months of life have been clearly shown to be responsible for the development of diabetes in NOD mice and BB rats. Recent human epidemiology data

10 shows that immunological events occurring at birth have a profound effect on the development of diabetes. These events include maternal fetal blood group incompatibility as well as exposure to rubella virus and nitrates at birth (Diabetes Reviews 1:15-42,1993; Diabetologia 35:671-765,1992). The concept of

15 diabetes in humans responding similarly to diabetes in NOD mice is widely accepted. Clinical trials have also shown that type I diabetes in humans can be prevented by immunosuppressants like cyclosporine when administered to prediabetics or newly diagnosed diabetics (Diabetes Reviews 1:15-42,1993). Immunosuppressants

20 have a similar effect on NOD mice and BB rats (Clinical and Investiga-tive Medicine 10:488-495,1987). The NIH recently embarked on a trial of screening up to 80,000 children to initiate a program of treating prediabetics with insulin immunotherapy, after a small phase I trial in humans supported

25 results of in NOD mice (Lancet 341:927-928,1993).

The correlation between the effects of a diabetes treatment on NOD mice, and its prospective effect in diabetic humans, is confirmed by Rossini, Mordes, and Like, Ann Rev. Immunolog. 3:289-320, 1985; Lampeter, Signore, Gale, and Pozzilli,

Diabetologia 32:703-708, 1989; Pacheo-Silva, Bastos, Muggia, Pankewez, Nichols, Murphy, Strom, and Rubbin-Kelley, Eur.J. Immunology 22:697-702,1992; Rossini, Handler, Greiner, and Mordes, Autoimmunity 18:221-225, 1991; Kolb, Diabetes/Metabolism

5 Reviews 3:751-758, 1987; Kikutani and Makino, Advances in Immunology 51: 285-321, 1992; Fox, J.Exp.Med. 175:1409-1412, 1992; Fitzpatrick, Lepault, Homo-Delarche, Bach, and Dardene, Endocrinology 129:1382-1390, 1991; Hawkins, Gala, and Dunbar P.S.E.B.M. 202:201-205, 1993; Jacob, Aiso, Michie, Mcdevitt,and

10 Acha-Obea; Proc. National Acad.Sci.USA 87:968-972, 1990; McInerney, Pek and Thomas, Diabetes 40:715-725, 1991; Karounos and Thomas, Diabetes 39:1085-1090, 1990; Yale and Marliss, Cln.Exp.Immunol 57:1-11, 1984; Kastern, Lang and Sorensen, Current Topics in Microbiology and Immunology 156:87-102, 1990;

15 Wang, Singh, Warnok, and Rajotte, Diabetes 41:114-117, 1992; Dotta and Eisenbarth, Clinical Immunology and Immunopathology 50:S85-S95, 1989; Elias, Reshef, Birk, van der Zee, Walker, Cohen, Proc. Natl. Acad. Sci. 88:3088-3091, 1991.

Many of the clinical characteristics of Type I diabetes in

20 humans and of the similar diabetes in NOD mice are found in BB rats, leading experts to believe diabetes in BB rats is also a autoimmune disorder. Insulitis develops in the pancreas of BB rats before the onset of diabetes while antibodies develop to islet cells and possibly to insulin. Diabetes can be prevented

25 by neonatal thymectomy as well as administration during the prediabetic period of cyclosporine, anti-lymphocyte antibodies, or purified lymphokines like TNF. Genetic experiments show that diabetes is closely linked to the MHC class II genes in BB rat as it is in humans. Many older rats develop autoimmune

thyroiditis that is casually related to the development of diabetes as occurs in humans. Several review papers have been published on this model (Diabetes/Metabolism Reviews 3:725-750,1987; Critical Reviews in Immunology 9:45-65,1989; Diabetes/Metabolism Reviews 8:9-37,1992).

In view of the fact that the methods of the present invention have, as shown by comparing examples 1, 2 and 4, been efficacious in two different species of mammal, we believe that persons of ordinary skill in the art would expect the invention to be adaptable, without undue experimentation, to humans. In this regard, we believe that those skilled in the art would find it significant that these parallel successes were achieved despite differences between diabetes in NOD mice and in BB rats. Diabetes develops in approximately equal numbers of males and females in contrast to NOD mice where disease develops more commonly in females. The incidence of diabetes in BB rats is not affected by gonadectomy or the administration of androgens as occurs with NOD mice. In contrast to humans and NOD mice, BB/Wor rats, the most commonly used substrain of BB rats, are severely lymphopenic. They have a marked decreased number of mature T lymphocytes in peripheral blood, spleen and lymph nodes. The CD4+ subset is substantially reduced but the CD8+ subset is almost completely absent. Natural killer cells are relatively over expressed. Several review papers have been published on this model (Mordes et al. 1987; Parfrey et al. 1989; Crisa et al. 1992).

Examples 3 and 5 show that the value of the invention is not limited to the prevention or treatment of diabetes, but rather is relevant to other immune-mediated disorders, too, such as

organ-specific   autoimmunity   (Ex.   3)   and   systemic   lupus
erythrematosis (SLE)(Ex. 5). Example 5 uses MRL/lpr mice as an
SLE model.

5   The MRL/lpr mice, as with human SLE patients, develop anti-
DNA and anti-nuclear autoantibodies which can cause immune
complexes. The immune complexes can cause causing arthritis,
dermatitis, and fatal immune complex glomerulonephritis if
untreated (J.Invest Dermatol 94:52-57, 1990).

The following are a few references verifying both the
10   similarity of the disease in MRL mice to SLE in humans and the
clinical importance of the MRL model. Berger, Perez, Laroche,
Edelson 1990, J. Investigative Dermatology 94:52-57; Halpern,
Hersh, Yocum 1990, Clinical Immunology and Immunopathology
55:242-254; Guitierrez-ramos, Andreu, Marcos, Vegazo, and
15   Martinez 1991, Autoimmunity 10:15-25; Shlomchick, Mascelli, Shan,
Radic et al. 1990; J.Exp. Medicine 171:265-297; Breneman,
Moynihan, Grota, Felten, Felten 1993; Brain, Behavior, & Immunity
7:135-43; Treadwell, Cohen, Williams, O'Brien, Volkman, Eisenberg
1993; Journal of Immunology 150:695-9; Lemire, Ince, Takashima
20   1992, Autoimmunity 12:143-8; Galli na, Steele 1991; Journal of
Autoimmunity, 4:755-68; Loor, Jachez, Montecino-Rodriguez, Klein,
Kuntz, et al. 1988; International Journal of Radiation Biology
& Related Studies in Physics, Chemistry & Medicine, 53:119-36;
Waterfield, Fairhurst, Chu, Levy, 1987; Immunology 61:173-8.

25   The MRL data is important not only because it is a good
model for human SLE but because this autoimmune disease is both
genetically, immunologically, and clinically very different from
diabetes.   Indeed, since the SLE-like disease spontaneously
developed by the MRL mice has a wide range of clinical and

serological characteristics that mimic not only human SLE but other autoimmune disorders such as Sjogren's syndrome, and rheumatoid arthritis (RA), see Bartlett, Popovic, Raiss 1988; Scandinavian Journal of Rheumatology - Supplement. 75:290-9, 1988, the MRL data can be deemed supportive of efficacy against these other autoimmune disorders, too.

EXAMPLE 1

The first Example involved injecting anthrax vaccine diluted in PBS into female non-obese diabetic prone (NOD) mice (n=19) (Taconic, Germantown, NY) on day 8 (.1 ml, 1:100), day 15 (.15 ml, 1:50), and day 29 (.2 ml, 1:50) of life. A second group of NOD (n=20) received plaque vaccine diluted in PBS on the same days but at a slightly lower dilution, day 8 (.1 ml, 1:50), day 15 (.15 ml, 1:50), and day 29 (.2 ml, 1:25). A third group of NOD (n=20) received a similar volume of PBS control on the same days as the mice in the first two groups.

Starting at approximately 16 weeks and continuing every 2 weeks until 28 weeks, tail blood was removed and checked for glucose using glucose sensitive chemstrips (Boehringer Mannheim, Indianapolis, IN). A blood glucose level over 300 mg/dl was considered positive. The cumulative incidence of diabetes in the anthrax treated group flattened out at 42.1% with no new cases detected after 24 weeks. The group receiving the plague vaccine appeared to begin flattening out and reached a cumulative incidence of 57.9% diabetic at 28 weeks. The PBS control group, showed a continual increase in the cumulative incidence of diabetes from 30% at 16 weeks to 65% at 28 weeks (Figure I). The remaining anthrax treated animals and PBS controls in experiment

1 were bled at 36 weeks. The net result was that no new cases
of diabetes were detected in the anthrax treated group from 24
to 36 weeks and the cumulative incidence of diabetes flattened
out at 42.1% compared to an incidence of 75% at 36 weeks in the

5 PBS treated animals (Figure 1). The flattening of cumulative
incidence of diabetes curve indicates that diabetes is prevented
not just delayed.

                              EXAMPLE 2

     A second set of experiments was designed to see if the

10 therapeutic potential of anthrax vaccine could be improved by
adding additional agents and changing the dosing schedule.
Anthrax vaccine was given alone, with the combined diphtheria
tetanus vaccine, or with the combined whole cell pertussis
diphtheria tetanus vaccine. A series of nine intraperitoneal

15 injections of the vaccines diluted in PBS were given to newborn
NOD female mice using the following protocol: day 1 (.1 ml,
1:100), day 3 (.1 ml, 1:100), day 10 (.15 ml, 1:100 ml), week 4
and every 2 weeks through week 14 (.2 ml, 1:50). Vaccines were
mixed prior to injection so volume in the notation refers to

20 total volume injected.

     Two control groups were used for Example 2. The first
control involved injecting PBS using the dosing protocol
mentioned above. The second control group in Example 2 received
a single intraperitoneal injection of whole cell DTP vaccine

25 diluted in PBS (.2 ml, 1:50) at 2 months of age. This group was
designed to see if the current immunization schedule of humans
could inhibit the development of diabetes.

The new dosing schedule used in Example 2 greatly reduced development of diabetes in the NOD mice. The group receiving anthrax vaccine alone (n=29) had a cumulative incidence of diabetes that peaked at 24 weeks and remained constant at 13.8% through the 32nd week. These results compare to a cumulative incidence of 42.1% in the group receiving anthrax vaccine in Example 1. The most remarkable results in Example 2 occurred with animals treated with anthrax vaccine combined with the whole cell pertussis, diphtheria and tetanus vaccine. None of 29 animals developed diabetes by 32 weeks of age (Figure 2).

The addition of the combined diphtheria and tetanus vaccine mixed with anthrax inhibited diabetes to a greater extent than anthrax alone but less than the combination of anthrax and DTP. Only 7.7% of animals receiving A + DT (n=26) developed diabetes by 32 weeks. A second group of mice (n=26) received a similar treatment of A + DT but were injected with DTP vaccine in place of DT at week 8. This was done to simulate the normal immunization of humans at 2 months with DTP. The animals in the later group developed a higher cumulative incidence of diabetes than those who did not receive pertussis component at 8 weeks, 23% versus 7.7% respectively at 32 weeks.

The injection of PBS slowed the progression of diabetes but the cumulative incidence curve did not flatten out during the 32 week study. The cumulative incidence in the PBS control was 48.1% at 32 weeks compared to 65% at 28 weeks in the PBS control of Example 1. The increased suppression of diabetes in the Example 2 group may be due to inflammation in the abdomen caused by the earlier and more frequent injections.

The control group receiving 1 injection of DTP at 8 weeks, developed a cumulative incidence of diabetes of 86.7% at 32 weeks. While the normal childhood vaccination schedule recommends a second injection at 16 weeks, 30% of animals already had diseases by this time and many of the rest probably had significant subclinical disease. A second injection would not have prevented disease in at least 30% of animals and thus the data clearly shows the current childhood immunization schedule for humans is ineffective in preventing immune mediated disorders.

EXAMPLE 3

Another experiment involved injecting newborn mice with 15 mg/kg/day of cyclosporine (Sandoz, East Hanover, NJ) intraperitoneally for the first 7 days of life to make the animals prone to developing autoimmunity (Sakaguchi and Sakaguchi 1989). Female C3H/Hen mice (Harlan Sprague Dawley, Indianapolis, IN) were then injected with a combination of the anthrax vaccine and the DT vaccine, or PBS control starting several days later to see if the vaccine would prevent the development of autoimmunity. The injection schedule was as follows: day 10 (.15 ml, 1:100), day 17 (.2 ml, 1:50) and every 2 weeks for 2 more injections (.2 ml, 1:50).

A second injection schedule was performed using anthrax vaccine mixed with DTP vaccine to determine if the effect derived from the first set of experiments could be magnified by including pertussis vaccine at higher doses. Injections were as follows: day 6-8 (0.15 ml, 1:10), day 14-16 (0.2 ml, 1:10) and Day 27-29

(0.2 ml, 1:10), where the notation 6-8 means one administration during the period of 6 to 8 days.

Tail blood was drawn from mice at the age of about 8 weeks and the resulting sera, diluted 1:80 in a solution containing PBS and 3% fetal calf serum, was screened for autoantibodies using ELISA assays against gastric antigens (Sakaguchi and Sakaguchi 1989). The microsomal antigens were plated on Immulon 3 (Dynatech Laboratories, Chantilly, VA). A second ELISA assay was prepared similarly by plating optimal amounts of E. coli DNA (Sigma, St. Louis, MO) on Immulon 3 plates. An alkaline phosphatase-conjugated anti-IgG Fc fraction (Jackson Immunoresearch, West Grove PA) was used as the secondary antibody and the substrate was a 2×10^{-4} M 4-Methyumbelliferyl phosphate solution (Classen and Shevach, 1991). Plates were read on a Dynatech MicroFLOUR machine which uses a 365 nm Broadband Filter for the excitation beam and 450 nm narrow band interference filter for the emission beam.

Of those receiving the ADT treatment 22% (n=23) were free of antigastric antibodies compared to 12% (n=25) of female mice which received the PBS control. One control animal had DNA antibodies compared to none of the ADT treated mice. The effect was significantly greater in those mice receiving anthrax vaccine and DTP, as 61% (n=23 males and females) did not develop antigastric antibodies. Methods and compositions of the present invention are thus effective in preventing or substantially reducing the incidence of several chronic immune mediated disorders.

EXAMPLE 4

Diabetic prone BB rats were immunized in order to show that the method of immunization could prevent diabetes in other species beside NOD mice. BB rats spontaneously develop diabetes at an early age as is the case in NOD mice and humans.

BB/Wor rats (U. of Massachusetts Medical School, Worchester MA,USA) were immunized with a combination of the anthrax and DTP vaccines (n=20) or nothing as a control (n=28). Groups contained approximately equal number of male and female rats. The vaccinated group was given the following dosing schedule: day 1 (.1ml,1:5); day 4 (.15ml, 1:5), day 11 (.15ml, 1:5), day 25 (.2ml, 1:5), day 39 (.2ml, 1:5), day 53 (.2ml, 1:5), day 61 (.2ml 1:2.5), and every 14 days for 3 more injections at approximately (.2ml, 1:2.5). Days of injection varied by one at times. (The notation 1:5 means 1 part vaccine to 5 parts PBS.)

At 16 weeks of age 54% of the untreated rats had developed diabetes and or died compared to 20% in the vaccinated group. At 20 weeks of age 54% of the untreated rats had developed diabetes and or died compared to 25% in the vaccinated group. At 32 weeks the results were 54% versus 35% respectively (Figure 3) which represents a 34% reduction in the incidence of diabetes. The difference between the two groups were statistically significant at 20 weeks (P=0.027).

The findings that the method of immunization can prevent diabetes in both NOD mice and BB rats provides strong proof that methods of immunization presented in the specifications have the ability to prevent chronic immune mediated diseases in mammals with very different genetic defects.

EXAMPLE 5

The methods of immunization taught herein also inhibit spontaneous autoimmunity in MRL/lpr mice, which develop a disease that closely resembles Systemic Lupus Erythematosus or SLE in humans.

5   Pregnant MRL/MpJ-lpr mice (Jackson Laboratory, Bar Harbor, Maine) gave birth to pups which were injected with PBS or a combination of the anthrax vaccine and the acellular DTP vaccine. A series of nine intraperitoneal injections of the vaccines, diluted in PBS, were given to newborn mice using the following

10   protocol: day 1 (.1 ml, 1:100), day 3 (.1 ml, 1:100), day 10 (.15 ml, 1:100 ml), week 4 and every 2 weeks through week 14 (.2 ml, 1:50). Weaning was done at approximately 21 days and only the female mice were used in the experiment. One group of 18 control mice received a similar injection schedule of PBS starting on day

15   1 and a second group of 20 control mice received a similar schedule (i.e., every 2 weeks through week 14) but starting at 4 weeks of age.

Urine was screened for the presence of protein, a common clinical test for glomerulonephritis. A few drops of mouse urine

20   were placed on a urine protein dipstick (Albustix, Miles Inc, Elkhart, IN). At 13 weeks 6/38 (15.8%)of PBS control mice had a urine with a protein over 300mg/dl while 0/28 of the vaccinated mice had a urine with a protein over 300mg/dl. At 14 weeks 8/38 (21%) of PBS control mice had a urine with protein of over

25   300mg/dl while 0/27 of the vaccinated mice had developed urines with similar protein levels. At 15 weeks 10/38 (26.3%)of PBS control mice had a urine with protein of over 300mg/dl while 2/26 (7.7%) of the vaccinated mice had developed urines with similar protein levels.

EXAMPLE 101

Epidemiology studies were performed to verify that immuniza-
tion practices with pharmaceutically acceptable doses of
pharmaceutically acceptable vaccines modulates the development
5   of diabetes. The data includes correlations between the specific
immunization schedules and the incidence of diabetes in western
European countries as well as temporal studies showing changes
in the incidence of diabetes in countries after the immunization
schedules were changed. Additional epidemiology data is provided
10 to explain the mechanism of action of this phenomenon and give
further support for the data showing vaccination can prevent the
development of immune mediated diabetes in humans.

1.   Intercountry Analysis.

15   Europe was chosen for an intercountry analysis because in
a relatively small geographic area there are many different
countries that have different immunization schedules and the
incidence of diabetes in the countries is known. The people in
the western European countries have closely related racial
20   backgrounds, diets, economic standards of living, and standards
of health care. Eastern European countries of the former
communist block were excluded because their standard of living
and standard of medical care is not up to western levels.

The study made the following assumptions: (1) the nations
25   included in the study had developed comparable quality of health
care allowing for similar risk of neonatal infections and similar
efficiency in diagnosing diabetes; (2) vaccines were of
comparable quality from country to country; (3) the amount of
diphtheria and tetanus toxoid given to people in different

countries did not induce a significant biological difference; and
(4) epidemics traveled freely across borders such that a person
in a given European state was as likely to be exposed to a given
pathogen as some one in another country.

5   The data correlating the incidence of diabetes to immuniza-
tion schedule in western European countries is presented in Table
I.   References pertaining to the immunization schedules and
incidence of diabetes of each country are included with table I.
Data on the incidence of diabetes was taken from the following
10   multiple sources.   (Green et al 1992 ; Nystrom, Dahlquist,
Rewers, Wall 1990; Helgason, Danielsen, and Thorsson 1992;
Metcalfe and Baum 1991; Levy-Marshal et al. 1990; Tuomilehto et
al 1991.) Immunization practices in each country were taken from
an assortment of publications.   Note that the incidence of
15   diabetes in Malta was significantly understated according to the
Schranz and Prikatsky, (Diabetic Medicine 6:228-231,1989) . The
authors calculated the incidence of diabetes in Malta using the
number of type I diabetics in public health clinics and the
number of children of a particular age in the country. The
20   authors admit significant number of diabetic children were
treated in private clinics that were not accounted for.

Sweden was one country that was difficult to characterize.
The country stopped immunizing newborns with BCG in the middle
of 1975 but the date it stopped revaccinating school children may
25   have been as late as 1986. Pertussis vaccination of newborns was
stopped in 1979.

The data in Table I indicates that administration of
vaccines after two months increases the incidence of diabetes
while administration of vaccines at birth can prevent diabetes.

The findings are highly statistically significant. Administration of the pertussis vaccine after two months of age explains the higher incidence of diabetes in group 3 compared to most regions in group 1. Administration of the BCG vaccine after two

5 months of age explains the higher incidence of diabetes in group 4 compared to group 3. Administration of the Hemophilus influenza vaccine after two months of age explains the higher incidence of diabetes in group 5 compared to 4. The ability of the BCG vaccine to protect against diabetes when administered at birth

10 explains the lower incidence of diabetes in group 2 compared to group 3. There appears to be a correlation between immunization rate and the incidence of diabetes within some groups since the districts in England and Ireland which had low immunization coverage with the pertussis vaccines compared to other countries

15 in their groups (World Health Statistics Annual, 1991 page 62) had the lowest incidence of diabetes in their groups. Luxembourg had the highest incidence of diabetes in group 3 which can be explained by a significant number of individuals receiving the BCG vaccine even though it is not required (ibid). Minor varia-

20 tions within a group can be explained by genetic variation, environmental differences including epidemics, use of additional vaccines and the percent of children fully immunized.

Epidemiology data in the United Kingdom supports the animal data that vaccination with DTP induces diabetes. During the

25 pertussis scare of the 70's and early 80's England had a significantly lower immunization rate for pertussis than Scotland. This can explain the lower incidence of diabetes in England than Scotland in 1988. Furthermore, in Scotland those from deprived neighborhoods received fewer vaccinations which

could explain the lower incidence of diabetes in areas of deprivation. (Metcalfe and Baum 1991; Patterson and Waugh 1992; Wrench, McWhirter, and Pearson 1991; Maclure and Stewart 1984).

The incidence of diabetes in Sardinia is the one major exception in the study showing that the incidence of diabetes correlates with diabetes. Sardinia has an very high incidence of diabetes that can not be explained by the routine pediatric immunization practices used on the island. The high incidence of diabetes however can be explained by the extremely high incidence of thalassemia on the island. It is estimated that as many as 1:250 live births in Sardinia during this time had beta thalassemia major (NY ACAD Science 612:215-225, 1990). Prevalence studies show that 16% of these patients older than 10 are diabetic (Archives of Diseases in Children 63:58-62, 1988). Prevalence studies tend to underestimate the frequency of diabetes, because some patients die, so the cumulative incidence of diabetes in patients homozygous for the thalassemia major gene in the 0-14 age group may be as high as 25 per 100 or 25%. The elevated incidence of diabetes in homozygotes can partially explain the increased incidence of diabetes in Sardinia. An elevated incidence of diabetes in heterozygotes for beta thalassemia or alpha thalassemia could explain the rest of the elevated incidence of diabetes. According to Cao et al. (Journal of Medical Genetics 15:443-447, 1978) about 12.5% of individuals living in Sardinia are heterozygous for beta thalassemia and many of these have disorders similar to those in the homozygotes though less severe.

2. Intracountry Temporal Analysis: Induction of Diabetes

Animal and human epidemiology data indicates that the incidence of diabetes will change when a country changes its immunization schedule. The data below suggests that if a vaccine is initially given after two months of age then the incidence of diabetes will go up. This situation was seen in both epidemiology studies of diabetes Finland and the United States.

A. Finland

In Finland the incidence of diabetes had been very stable in the 0-4 year old age group from 1966-1975 until after the government made several changes to its vaccination schedule which was followed by large increases in the incidence of diabetes in the years 1976-1978 and 1988-1990 (Diabetologia 36:1303-1308, 1993). The rises in the incidence of diabetes can be explained by the addition of three new vaccines to the Finnish vaccine schedule. A large clinical trial immunizing 130,000 infants aged three months to 5 years old with Hemophilus influenza or menin-gococci polysaccharide vaccines was started in November, 1974 (New England Journal Medicine 297:686-691, 1977). Finland has a small population with less than 65,000 children born each year (World Health Statistics Annul) so 130,000 represented a signifi-cant proportion of children in the country 0-4 years of age. In 1976 the pertussis vaccine in Finland was changed by the addition of a second strain of bacteria making the vaccine more antigenic (Acta Paediatric Scand 298 Suppl 21-25, 1982). The vaccine regiment was next altered to include the measles, mumps, rubella, (MMR) vaccine in 1982. A trial of Hemophilus influenza conjugate vaccine was initiated in January 1986 and included 114,000 children born between October 1,1985 and August 31,1987. Based

on the results of the trial the Hemophilus influenza vaccine became part of the standard vaccine schedule in Finland starting in January of 1988 (New England Journal Medicine 317:1381-1387, 1990).

5   The changes in the incidence of diabetes especially in the 0-4 year old age group in Finland correspond temporally very closely with the changes in the vaccine schedule, supporting the rodent and the intercountry epidemiology data that the development of immune mediated diabetes is influenced by vaccination 10 (Table II). The use of the Hemophilus influenza vaccine as well as the addition of a second strain of bacteria to the pertussis vaccine explains the rapid rise of diabetes seen in the years 1977-1979 in 0-4 year olds. The discontinuation of the Hemophilus influenza vaccine can explain the drop in the incidence of 15 diabetes in 0-4 year olds from 19.3 in the 1977-1979 cohort to 16.0 in the 1980-1982 cohort (Diabetes Care 8, Suppl 1:10-16, 1985). The widespread use of the MMR vaccine starting in 1982 and the Hemophilus influenza vaccine in 1986 can explain the large rise in the incidence of diabetes in the 0-4 age group between 20 the years 1980-1982 and 1987-1989 (Diabetologia 36:1303-1308, 1993).   .

Temporal data comparing the incidence of diabetes in different age groups suggest that the use of more antigenic vaccines schedules is causing the increased incidence of diabetes 25 in Finland since the increased incidence is only seen in those who received the new vaccines.   The temporal rises in the incidence of diabetes in the 0-4 year old age group occur shortly after the new vaccines are added but rises in the incidence of diabetes in the older age groups are delayed until those children

immunized with the new vaccine reach the older age group. For example, following the use of the Hemophilus influenza/mening-ococcus vaccine in 1974 and new pertussis vaccine in 1976 the incidence of diabetes in the 1977-1979 cohorts increased compared

5   to 1970-1976 cohorts by 63.5% in 0-4 year olds, 16% in 5-9 year olds, 0% in 10-14 year olds (Table II). Changes in incidence of diabetes were statistically more significant in the 0-4 age group, the age group that received the vaccine. When the incidence of diabetes started to decline in 0-4 year olds in

10 1980-1982 following discontinuation of the Hemophilus influenza/meningococci vaccine, the incidence continued to rise in 5-9 and 10-14 year olds as those who received the Hemophilus influenza/meningococci vaccine and the new pertussis vaccine entered these age groups.

15 B. Allegheny County,Pennsylvania (Greater Pittsburgh area)

      Changes in the incidence of diabetes in Allegheny County, Pennsylvania (Diabetes Care 16:1606-1611 1993) can also be explained by changes in immunization schedules, supporting findings discussed earlier that the incidence of immune mediated

20 diabetes in humans is affected by vaccination. The epidemic of diabetes occurring in the 0-4 age group during the years 1985-1989, (Table III) can be explained by the addition of the Hemophilus influenza vaccine to the immunization schedule. The FDA approved the Hemophilus influenza polysaccharide vaccine in

25 1985 and the conjugated vaccine in 1987 (Jama 269:221-226, 1993). The vaccine was widely administered to children in Allegheny County. A study of its efficacy performed in Allegheny county showed that about 36% of children, chosen as controls, were

immunized with the vaccine between August, 1985 and July, 1987 (Jama 260:1419-1422, 1988). Based on epidemiology and animal data discussed earlier it was predicted that the incidence of diabetes would rise in Allegheny County after wide spread use of the Hemophilus influenza vaccine occurred.

The drop in the incidence of diabetes in the 0-4 age group in years 1975-1979 can be explained by a drop in pertussis immunization. In September, 1974, at a time when there was international concern about the incidence of encephalitis caused by the pertussis vaccine, the Pennsylvania state government passed Act 210 of 1974 requiring children to receive certain immunizations prior to entering school (Pennsylvania Medicine December 1974, p41-42). The law required immunization with polio, diphtheria, tetanus, measles and rubella, but not with pertussis vaccine. The law was a clear message that immunization with the pertussis vaccine was not considered necessary. Following the passage of this law, incidence of diabetes in the 0-4 age group dropped about 61% in the 1975-1979 time period as compared to 1965-1969 (P<.001) as shown in Table III. During the period of 1975 to 1979, immunization with the pertussis vaccine dropped of in several countries. Immunization with the pertussis vaccine was stopped in Sweden and pertussis vaccine acceptance in the United Kingdom fell from 75% in 1974 to 30% in 1978. (Pediatric Infectious Disease Journal 6:364-371, 1987).

The incidence of diabetes in the 0-4 age group during the period 1980-1984 rose to levels even higher than before 1975. This can be explained by the Pennsylvania Department of Health calling for increased immunization with the pertussis vaccine following an epidemic of pertussis in Pennsylvania in 1982

(Pennsylvania Medicine, March, 1983, p. 16) and new state laws requiring that children receive the mumps vaccine (Pennsylvania Medicine, January, 1983, pp. 12-16). The results are consistent with previous findings that changes in immunization policy can
5   lead to changes in the incidence of immune mediated diabetes.

3.Intracountry Temporal Analysis; Prevention of Diabetes

Epidemiology data from the Netherlands provides evidence that immunization with the smallpox vaccine at birth can prevent
10 the development of diabetes. This data supports the intercountry epidemiology data that immunization at birth with the BCG vaccine can prevent diabetes. The findings also support the temporal data presented above that changes in immunization practices can effect the development of immune mediated diabetes in humans.
15 Epidemiology data shows that in the cumulative incidence of diabetes up to the age of 18 differed significantly in Danish birth cohorts (Diabetologia 35:139-142, 1992). There were two significant drops in the incidence of diabetes; one was centered around 1962 when the cumulative incidence dropped to 1.1 per 1000
20 ($P<.05$), and the other was centered around 1966 when the cumula-tive incidence dropped to 1.71 per 1000. The drops are in contrast to a cumulative incidence of diabetes outside of these troughs of about 1.98 per 1000 (Table IV). The drops in 1962 and 1966 both occurred during smallpox epidemics in Europe and can
25 best be explained by immunization of newborn infants in these periods with smallpox vaccines.

Western Europe had major epidemics of smallpox centered around the years 1962 and 1966. The epidemic in 1962 actually started in 1961 continuing into 1963 and included 158 cases from

seven western European countries. The 1966 epidemic included 72
cases and was limited to the United Kingdom. There was a strong
emphasis placed on vaccination in smallpox epidemic of 1961-1963
as demonstrated by World Health Organization statistics showing
5   23.5 million Europeans were vaccinated with the smallpox vaccine
in 1962 compared to an norm of about 11 million in nonepidemic
years   (International   Symposium   on   Vaccination   against
communicable diseases Monaco 1973; Symp. Series Immunobiol.
Standard 22:13-24.) Changes in smallpox vaccine acceptance were
10   detected around 1962 and 1966 in the Netherlands (ibid pp. 271-
276).

Literature from the 1960s and earlier suggested humans could
be immunized at any time after birth, but immunization before
three months of age was associated with an increased risk of
15   encephalitis.   It was customary at the time for physicians
practicing in areas with a low incidence of smallpox to wait
until the patient was several months old before administering the
smallpox vaccine, however in areas with high incidence of
smallpox, like third-world countries, smallpox vaccine was often
20   given at birth (International Symposium on Smallpox Vaccine,
Bilthoven 1972. Symp. Series Immunobiol. Standard 19:243-248;
Tubercle 43:155-160, 1962).

The common practice in the Netherlands in the 1960s was to
immunize children with the smallpox vaccine starting at two
25   months of age in normal, nonepidemic conditions (International
Symposium on Smallpox Vaccine, Bilthoven 1972. Symp.   Series
Immunobiol. Standard 19:235-242).   Given the fact that the
literature recommends immunization earlier than usual in times
of epidemics, it would have been expected that a number of

physicians would have given the vaccine several weeks earlier as in four weeks of age or at birth. The resulting switch in immunization explains the drop in the incidence of diabetes in the cohorts born during smallpox epidemics.

5   **4.Related epidemiology data**

Epidemiology data suggests that diabetes in humans can be caused by transient immunological changes at birth which gives further support to the finding that immunization can alter the development of diabetes in humans. A report has been published
10   that maternal-child blood group incompatibility is associated with an increased risk for developing diabetes (OR =1.61) (Diabetologia 1992; 35:671-675). The more severe cases of the disorder had a higher incidence of the disease. Maternal-child blood group incompatibility has been shown by others (New England
15   Journal of Medicine August 22, 1974, p.420) to cause transient suppression of the thymus/T cell immune system. These findings demonstrate that in humans, as in rodents, transient alterations to the developing immune system, immunosuppression or stimula-tion, will alter the risk of developing immune mediated diabetes
20   or another chronic immune mediated disease later in life, enhance or decrease respectively.

All references cited herein, including journal articles or abstracts, published or corresponding U.S. or foreign patent applications, issued U.S. or foreign patents, or any other
25   *references, are entirely incorporated by reference herein, including all data, tables, figures, and text presented in the cited references. Additionally, the entire contents of the*

references cited within the references cited herein are also entirely incorporated by reference.

Reference to known method steps, conventional methods steps, known methods or conventional methods is not in any way an admission that any aspect, description or embodiment of the present invention is disclosed, taught or suggested in the relevant art.

The foregoing description of the specific embodiments will so fully reveal the general nature of the invention that others can, by applying knowledge within the skill of the art (including the contents of the references cited herein), readily modify and/or adapt for various applications such specific embodiments, without undue experimentation, without departing from the general concept of the present invention. Therefore, such adaptations and modifications are intended to be within the meaning and range of equivalents of the disclosed embodiments, based on the teaching and guidance presented herein. It is to be understood that the phraseology or terminology herein is for the purpose of description and not of limitation, such that the terminology or phraseology of the present specification is to be interpreted by the skilled artisan in light of the teachings and guidance presented herein, in combination with the knowledge of one of ordinary skill in the art.

Any description of a class or range as being useful or preferred in the practice of the invention shall be deemed a description of any subclass or subrange contained therein, as well as a separate description of each individual member or value in said class or range.

TABLE I: Intercountry Analysis

Immunization Schedule Versus Incidence of Type I Diabetes

Group	Immunization Schedule	Year	Incidence of Diabetes (Mean)	P Values	
1	No Pertussis, No BCG		(14.5)		
	Italy, Lombardi(6,13,1)	1988	6.8		
	Italy, Lazio(6,13,1)	1987	6.5		
	Italy, Sardinia(6,13,1)	1987	30.2		
2	Pertussis, BCG before 2 months		(7.45)	X	
	R. of Ireland (6,2)	1988	6.8		
	France (6,1)	1990	7.8		
	Austria (6,1)	1989	7.7		
	Portugal (6,1)	1986	7.5		
3	Pertussis, No BCG		(10.92)	<0.02	X
	Iceland (7,3)	1980-1989	10.8		
	Netherlands (6,1)	1989	11		
	Spain, Catelinia (6,1)	1986	10.6		
	Spain, Madrid (6,16)	1985-1988	10.9		
	Belgium (8,1)	1989	9.8		
	Luxembourg (6,1)	1989	12.4		
4	Pertussis, BCG Vaccination School Age		(20.01)	<.0001 <.005	X
	England, Oxford (6,1)	1988	16.4		
	Northern Ireland (6,2)	1988	16.6		
	Scotland (6,2)	1988	19.8		
	Denmark (6,12,1)	1990	21.5		
	Norway (15,1)	1989	20.8		
	Malta (9,4)	1980-1987	(13.6+)		
	Sweden (6,10,11,5)	1987	25		

5 Pertussis, Hib, BCG Vaccination at 3 months and School

                                    (42.9)   <.0001 <.0001 <.0001
       Finland (6,14,1)   1988   42.9

Incidences of type I diabetes mellitus are reported as yearly figures per 100,000 children as recorded in the references indicated. Statistics were generated from unweighted group means using a normal approximation to the poisson distribution. All statistics shown where highly significant using T=1 with the exception of group 3 where (P<0.02) was calculated using T=4. Sardinia was the only major exception to the trend as discussed in the text. The incidence of diabetes in Malta was significantly underestimated as discussed in reference 4. The immunization schedule of Sweden has changed during the 15 years prior to 1987. Most notably the pertussis vaccination was discontinued in 1979.

References: (1) Lancet 339:905-909, 1992 (2) British Medical Journal 302:443-447, 1991 (3) Diabetologia 35:880-883, 1992 (4) Diabetic Medicine 6:228-231, 1989 (5) International Journal of Epidemiology 19:141-146, 1990 (6) World Health Organization, Working Group on Immunization of Tourists and other Travelers Venice, November 1990, By B. Bytchenko (7) Vaccination policy and status in Iceland, Iceland Government (8) Symp. Series Immunobiol. Standard, 22:223-225, 1973 (9) Personal Communication Malta pediatrician, #011-356-241-251 (10) Pediatr. Infect. Dis J. 6:364-371, 1987 (11) Personal Communication Dr. Ingrid Trolin, Uppsala Sweden #46 18 17 46 61 (12) Symp. Series Immunobiol. Standard, 22:235-237, 1973 (13) Pediatr. Infect. Dis. J. 11:653-661,1992 (14) N.Engl.J.Med 323:1381-1387, 1990 (15) Tubercle 69:119-123, 1988 (16) Diabetologia 33:422-424, 1990.

Table II

Incidence of Type I Diabetes in Finland

Age Group	Years	N	Incidence of Diabetes	% Change Incidence	P values	
0-4	1970-76	262	11.8		X	
	1977-79	174	19.3	63.6%	P<0.001	
	1980-82	152	16	-17.1%	P<0.005	X
	1987-89	243	25.9	61.9%	P<0.0001	P<0.0001
5-9	1970-76	729	27.6		X	
	1977-79	304	32	15.9%	P<0.05	
	1980-82	299	33	3.1%	P<0.02	X
	1987-89	382	39.3	19.0%	P<0.0001	P<0.05
10-14	1970-76	1047	37.5		X	
	1977-79	421	37.5	0%	N.S.	
	1980-82	399	38.6	2.93%	N.S.	X
	1987-89	389	40.6	5.2%	N.S.	N.S.

Incidences of type I diabetes mellitus are reported as yearly figures per 100,000 children as recorded in the references indicated. The per cent change in incidence was calculated using the incidence in the prior year as the reference. Statistics were recorded as reported in the references when available. Statistics not reported were calculated using a normal approximation to the poisson distribution. Differences that were not statistically significant are marked N.S.

References

Diabetologia 35:70-76, 1992

Diabetes Care 8 Suppl 1:10-16, 1985

Diabetologia 36:1303-1308, 1993

Table III

Incidence of Type I Diabetes Mellitus

Age 0-4, Allegheny County, Pennsylvania

Years	Incidence of Diabetes	% Change Incidence	P Values
1965-1969	6.75	X	
1970-1974	6.375	-5.55%	N.S.
1975-1979	2.625	-58.82%	P<.001
1980-1984	9.875	276.19%	P<.02
1985-1989	16.125	63.29%	P<.0001

Total # of cases 183

Incidences of type I diabetes mellitus are reported as yearly figures per 100,000 children as recorded in the reference indicated. The per cent change in the incidence was calculated using the incidence in the prior year as the reference. Statistics were calculated using a normal approximation to the poisson distribution. The results show a highly statistically significant decline in the incidence of type I diabetes mellitus in the years 1975-1979 as well as statistically significant rise during the years 1980-1989. Differences that were not statistically significant were marked N.S.

Reference

Diabetes Care 16:1606-1611,1993

Table IV

Cumulative Incidence of Type I Diabetes Mellitus

Danish Military Recruits Age 18

Year Born	# of Diabetics	Cumulative Incidence Diabetes/1000	P value	Smallpox Cases Western Europe
1960	189	1.85		1
1961	167	1.76		79
1962	96	1.11	P<0.05	51
1963	188	1.5		28
1964	198	1.83		0
1965	203	1.93		1
1966	184	1.71	N.S.	72
1967	219	2.07		2
1968	194	1.96		2
1969	264	2.11		0
1970	234	2.12		22

Cumulative incidences of type I diabetes mellitus in Danish military recruits under 19 years of age are reported per 1000 recruits as recorded in the reference indicated. Statistics were calculated by the authors in the reference. The results show a statistically significant decline in the incidence of type I diabetes mellitus in the years 1961-1963 which corresponded to a smallpox epidemic. The decline in the incidence of diabetes in 1962 followed the epidemic of 1961. The incidence of diabetes also declined in 1966 during a second smallpox epidemic. The later was not statistically significant, N.S.

Reference

Diabetologia 35:139-142, 1992

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Table V

Potential Immunization Schedules

	Week	Schedule 1	Schedule 2
5	0	DTP,Hib,HepB,IPV,MMR,NPI	
	1		
	2	DTP,Hib,HepB,IPV,MMR,NPI	DTP,Hib,HepB,OPV,MMR
	3		
	4	DTP,Hib,HepB,IPV,MMR,NPI	DTP,Hib,HepB,OPV,MMR
10	5		
	6	DTP,Hib,HepB,IPV,MMR,NPI	DTP,Hib,HepB,OPV,MMR
	7		
	8	DTP,Hib,HepB,IPV,MMR,NPI	DTP,Hib,HepB,OPV,MMR
	9		
15	10	DTP,Hib,HepB,IPV,MMR,NPI	DTP,Hib,HepB,OPV,MMR
	11		
	12	DTP,Hib,HepB,IPV,MMR,NPI	DTP,Hib,HepB,OPV,MMR
	13		
	14	DTP,Hib,HepB,IPV,MMR,NPI	DTP,Hib,HepB,OPV,MMR
20	15		
	16	DTP,Hib,HepB,IPV,MMR,NPI	DTP,Hib,HepB,OPV,MMR
	17		
	18	DTP,Hib,HepB,IPV,MMR,NPI	DTP,Hib,HepB,OPV,MMR

	Week	Schedule 3	Schedule 4
25	0	DTP,Hib,HepB,IPV,MMR,NPI	DT,Hib,HepB,IPV
	1		
	2		DTP,Hib,HepB,IPV,MMR
	3	DTP,Hib,HepB,IPV,MMR,NPI	
30	4		DTP,Hib,HepB,IPV,MMR
	5		
	6	DTP,Hib,HepB,IPV,MMR,NPI	DTP,Hib,HepB,IPV,MMR
	7		
	8		

9	DTP,Hib,HepB,IPV,MMR,NPI	DTP,Hib,HepB,IPV,MMR
10		
11		
12	DTP,Hib,HepB,IPV,MMR	DTP,Hib,HepB,IPV,MMR
13		
14		
15	DTP,Hib,IPV,MMR	DTP,Hib,HepB,IPV,MMR
16		
17		
18	DTP,Hib,IPV,MMR	DTP,Hib,HepB,IPV,MMR

The four immunization schedules above are nonlimiting examples of immunization schedules that can be used to reduce the incidence of diabetes as compared to standard immunization schedules listed in the specifications. The following abbreviations are used above: DTP - diptheria, tetanus, pertussis; Hib-Hemophilus influenza B; HepB hepatis B; IPV -inactivated polio virus; OPV - oral polio virus, MMR - measles mumps rubella, NPI - nonpediatric immunogen.

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I claim:

1. A method of determining whether an immunization schedule affects the incidence, prevalence, frequency or severity of a chronic immune-mediated disorder in at least one treatment group of mammals, relative to a control group of mammals, which comprises immunizing mammals in at least one treatment group of mammals with one or more doses of one or more immunogens, according to said immunization schedule, and comparing the incidence, prevalence or frequency of said chronic immune-mediated disorder in the treatment group, with that in the control group;

where said chronic immune-mediated disorder is diabetes, and said mammals are human;

wherein the control and treatment group differ by at least one of the following differences:

  a)   the presence of at least one immunogen in the schedule for one group and not the other;
  b)   a difference in the size of the dose of at least one immunogen administered to both groups;
  c)   a difference in the number of doses of at least one immunogen administered to both groups; or
  d)   a difference in the day of administration, relative to birth, of the first dose of at least one immunogen administered to both groups; and

wherein the effect of the schedule on the incidence, prevalence, or frequency of the disorder is observed at least one year after the first difference in immunization between the groups is manifest.

2. The method of claim 1 where at least one of said differences (a)-(d) relates to at least one immunogen other than a BCG immunogen.

3. The method of claim 1 where at least one of said differences (a)-(d) relates to at least one immunogen other than a BCG or measles immunogen.

4.   The method of claim 1 where at least one of said differences (a)-(d) relates to at least one immunogen other than a BCG, measles, mumps, rubella, smallpox, diphtheria, tetanus, pertussis or polio immunogen.

5.   The method of claim 3 where at least difference (a) applies.

6.   The method of claim 3 where at least difference (b) applies.

7.   The method of claim 3 where at least difference (c) applies.

8.   The method of claim 3 where at least difference (d) applies.

9.   The method of claim 3 where at least two of differences (a)-(d) apply.

10.   The method of claim 3 where at least three of differences (a)-(d) apply.

11.   The method of claim 1 where all of differences (a)-(d) apply.

12.   The method of claim 1 which further comprises determining whether an immunization schedule slows the onset of diabetes as evidenced by a reduction of the incidence of diabetes in one of said groups relative to another of said groups, as determined at a particular time or times after birth.

13.   The method of claim 1, where at least one observation of the effect of the schedules is made at least three years after said first difference in immunization of said mammals.

14.   The method of claim 1, where at least one of the following conditions applies

i) the first dose of said immunization schedule is given in at least one group when the mammals are less than 42

days old,

ii) said immunization schedule protects against at least two infectious diseases,

iii) the ability of said immunogen or immunization schedule to prevent an infectious disease is also tested,

iv) the treatment and control groups differ as to the presence, dosage, timing or composition of at least one immunogen which is other than a live vaccine,

v) said immunogen is one other than a pertussis immunogen,

vi) the method is part of a development process or clinical trial of a vaccine to test a vaccine for safety or efficacy,

vii) the method is prospective,

viii) said mammals are randomized in said treatment and control groups,

ix) at least one said treatment group receives one potentially pharmaceutically acceptable dose of at least two potentially pharmaceutically acceptable immunogenic agents which comprise at least one potentially pharmaceutically acceptable first pediatric immunogen and at least one agent selected from the group consisting of a second pediatric immunogen and a non-pediatric immunogen,

x) at least one immunogen being studied for its effect on the incidence or severity of said disorder is administered starting after 41 days but before 180 days of life,

xi) at least the majority of the mammals in the control group did not develop the infectious diseases which are prevented by said immunogen,

xii) mammals are excluded from a treatment group if at

least one of conditions (aa)-(cc) applies:

    aa) said mammals have substantial immunologic protection against the infectious disease which said immunization schedule protects against, or

    bb) said mammals have substantial levels of at least one surrogate marker of an autoimmune disease even though the mammals had not been previously diagnosed as having an autoimmune disease, or

    cc) said surrogate marker was substantially increased following a previous vaccination, infection or other immunologic challenge

xiii) at least one observation of the effect of the schedules occurs at least five years after the birth of the mammals

xiv) at least one chronic immune-mediated disorder in addition to diabetes is compared,

xv) at least one immunogen administered to the treatment group is selected from the group consisting of BCG, measles, mumps, rubella, diphtheria, pertussis, hemophilus influenza, tetanus, hepatitis B, polio, anthrax, plague, encephalitis, meningococcal, meningitis, pneumococcus, pneumonia, typhus, typhoid fever, streptococcus, staphylococcus, neisseria, lyme disease, cholera, E. coli, shigella, leishmania, leprosy, cytomegalovirus (CMV), respiratory syncytial virus, Epstein Barr virus, herpes, influenza, parainfluenza, rotavirus, adenovirus, human immunodeficiency virus (HIV), hepatitis A, NonA NonB hepatitis, varicella, rabies, yellow fever, Japanese encephalitis, flavirus, dengue, toxoplasmosis, coccidiomycosis, schistosomiasis, and malaria immunogens

xvi) it is determined whether the age of the subject mammal, at the time of commencement of the immunization schedule, affects the incidence, prevalence or frequency of the disorder,

xvii) the incidence of the disorder in the treatment group is compared to the incidence of the disorder in the control group, or

xviii) in at least one group both a pediatric immunogen and a non-pediatric immunogen are administered.

15. The method of claim 14 where condition (i), condition (xv) and at least one of the conditions (ii)-(xiv) and (xvi-(xviii) applies.

16. The method of claim 15 where at least two of conditions (ii)-(xiv) and (xvi)-(xviii) apply.

17. The method of claim 15 where at least three of conditions (ii)-(xiv) and (xvi)-(xviii) apply.

18. The method of claim 15 where at least four of conditions (ii)-(xiv) and (xvi)-(xviii) apply.

19. The method of claim 15 where at least five of conditions (ii)-(xiv) and (xvi)-(xviii) apply.

20. The method of claim 15 where at least six of conditions (ii)-(xiv) and (xvi)-(xviii) apply.

21. The method of claim 15 where at least seven of conditions (ii)-(xiv) and (xvi)-(xviii) apply.

22. The method of claim 15 where at least eight of conditions (ii)-(xiv) and (xvi)-(xviii) apply.

23. The method of claim 15 where at least nine of conditions (ii)-(xiv) and (xvi)-(xviii) apply.

24. The method of claim 15 where at least ten of conditions (ii)-(xiv) and (xvi)-(xviii) apply.

25. The method of claim 15 where at least eleven of conditions (ii)-(xiv) and (xvi)-(xviii) apply.

26. The method of claim 15 where at least twelve of

conditions (ii)-(xiv) and (xvi)-(xviii) apply.

27. The method of claim 15 where at least thirteen of conditions (ii)-(xiv) and (xvi)-(xviii) apply.

28. The method of claim 14 where conditions (x) and (xv), and at least one of conditions (i)-(ix), (xi)-(xiv) and (xvi)-(xviii) applies.

29. The method of claim 28 where at least two of conditions (i)-(ix), (xi)-(xiv) and (xvi)-(xviii) apply.

30. The method of claim 28 where at least three of conditions (i)-(ix), (xi)-(xiv) and (xvi)-(xviii) apply.

31. The method of claim 28 where at least four of conditions (i)-(ix), (xi)-(xiv) and (xvi)-(xviii) apply.

32. The method of claim 28 where at least five of conditions (i)-(ix), (xi)-(xiv) and (xvi)-(xviii) apply.

33. The method of claim 28 where at least six of conditions (i)-(ix), (xi)-(xiv) and (xvi)-(xviii) apply.

34. The method of claim 28 where at least seven of conditions (i)-(ix), (xi)-(xiv) and (xvi)-(xviii) apply.

35. The method of claim 28 where at least eight of conditions (i)-(ix), (xi)-(xiv) and (xvi)-(xviii) apply.

36. The method of claim 28 where at least nine of conditions (i)-(ix), (xi)-(xiv) and (xvi)-(xviii) apply.

37. The method of claim 28 where at least ten of conditions (i)-(ix), (xi)-(xiv) and (xvi)-(xviii) apply.

38. The method of claim 28 where at least eleven of conditions (i)-(ix), (xi)-(xiv) and (xvi)-(xviii) apply.

39. The method of claim 28 where at least twelve of conditions (i)-(ix), (xi)-(xiv) and (xvi)-(xviii) apply.

40. The method of claim 28 where at least thirteen of

conditions (i)-(ix), (xi)-(xiv) and (xvi)-(xviii) apply.

41. The method of claim 28 where at least fourteen of conditions (i)-(ix), (xi)-(xiv) and (xvi)-(xviii) apply.

42. The method of claim 28 where at least fifteen of conditions (i)-(ix), (xi)-(xiv) and (xvi)-(xviii) apply.

43. The method of claim 28 where all sixteen of conditions (i)-(ix), (xi)-(xiv) and (xvi)-(xviii) apply.

44. The method of claim 3 where the following condition applies

   i) the first dose of said immunization schedule is given in at least one group when the mammals are less than 42 days old.

45. The method of claim 3 where the following condition applies

   ii) said immunization schedule prevents at least one infectious disease.

46. The method of claim 3 where the following condition applies

   iii) the ability of said immunogen or immunization schedule to prevent an infectious disease is also tested.

47. The method of claim 3 where the following condition applies

   iv) the treatment and control groups differ as to the presence, dosage, timing or composition of at least one immunogen which is other than a live vaccine.

48. The method of claim 3 where the following condition applies

   v) said immunogen is one other than a pertussis immunogen.

49. The method of claim 3 where the following condition applies

vi) the method is part of a development process or clinical trial of a vaccine to test a vaccine for safety or efficacy.

50. The method of claim 3 where the following condition applies

vii) the method is prospective.

51. The method of claim 3 where the following condition applies

viii) said mammals are randomized in said treatment and control groups.

52. The method of claim 3 where the following condition applies

ix) at least one said treatment group receives one potentially pharmaceutically acceptable dose of at least two potentially pharmaceutically acceptable immunogenic agents which comprise at least one potentially pharmaceutically acceptable first pediatric immunogen and at least one agent selected from the group consisting of a second pediatric immunogen and a non-pediatric immunogen.

53. The method of claim 3 where the following condition applies

x) at least one immunogen being studied for its effect on the incidence or severity of said disorder is administered starting after 41 days but before 180 days of life.

54. The method of claim 3 where the following condition applies

xi) at least the majority of the mammals in the control group did not develop the infectious diseases which are

prevented by said immunogen.

55. The method of claim 3 where the following condition applies

xii) mammals are excluded from a treatment group if at least one of conditions (aa)-(cc) applies:

aa) said mammals have substantial immunologic protection against the infectious disease which said immunization schedule protects against, or

bb) said mammals have substantial levels of at least one surrogate marker of an autoimmune disease even though the mammals had not been previously diagnosed as having an autoimmune disease, or

cc) said surrogate marker was substantially increased following a previous vaccination, infection or other immunologic challenge.

56. The method of claim 3 where the following condition applies

xiii) at least one observation of the effect of the schedules occurs at least five years after the birth of the mammals.

57. The method of claim 3 where the following condition applies

xiv) at least one chronic immune-mediated disorder in addition to diabetes is compared.

58. The method of claim 3 where the following condition applies

xv) at least one immunogen administered to the treatment group is selected from the group consisting of BCG, measles, mumps, rubella, diphtheria, pertussis, hemophilus influenza, tetanus, hepatitis B, polio, anthrax, plague, encephalitis, meningococcal, meningitis,

pneumococcus, pneumonia, typhus, typhoid fever,
streptococcus, staphylococcus, neisseria, lyme disease,
cholera, E. coli, shigella, leishmania, leprosy,
cytomegalovirus (CMV), respiratory syncytial virus,
Epstein Barr virus, herpes, influenza, parainfluenza,
rotavirus, adenovirus, human immunodeficiency virus
(HIV), hepatitis A, NonA NonB hepatitis, varicella,
rabies, yellow fever, Japanese encephalitis, flavirus,
dengue, toxoplasmosis, coccidiomycosis, schistosomiasis,
and malaria immunogens.

59. The method of claim 3 where the following condition
applies

xvi) it is determined whether the age of the subject
mammal, at the time of commencement of the immunization
schedule, affects the incidence, prevalence or frequency
of the disorder.

60. The method of claim 3 where the following condition
applies

xvii) the incidence of the disorder in the treatment
group is compared to the incidence of the disorder in the
control group.

61. The method of claim 3 where the following condition
applies

xviii) in at least one group both a pediatric immunogen
and a non-pediatric immunogen are administered.

62. The method of claim 3 where at least one of the
following conditions applies:

i) the first dose of said immunization schedule is given
when the mammals are less than 42 days old,

ii) said immunization schedule prevents at least one
infectious disease,

iii) the ability of said immunogen or immunization
schedule to prevent an infectious disease is also tested,

iv) the treatment and control groups differ as to the presence, dosage, timing or composition of at least one immunogen which is other than a live vaccine.

63.   The method of claim 62 where at least two of conditions (i)-(iv) apply.

64.   The method of claim 62 where at least three of conditions (i)-(iv) apply.

65.   The method of claim 62 where all four of conditions (i)-(iv) apply.

66.   The method of claim 1 where the following conditions apply:

iii) the ability of said immunogen or immunization schedule to prevent an infectious disease is also tested,

vii) the method is prospective,

viii) said mammals are randomized in said treatment and control groups,

xiv) at least one chronic immune-mediated disorder in addition to diabetes is compared.

67.   The method of claim 66 where the following condition applies:

i) the first dose of said immunization schedule is given when the mammals are less than 42 days old.

68.   The method of claim 67 where the following condition applies:

xv) at least one immunogen administered to the treatment group is selected from the group consisting of BCG, measles, mumps, rubella, diphtheria, pertussis, hemophilus influenza, tetanus, hepatitis B, polio, anthrax, plague, encephalitis, meningococcal, meningitis, pneumococcus, pneumonia, typhus, typhoid fever, streptococcus, staphylococcus, neisseria, lyme disease,

cholera, E. coli, shigella, leishmania, leprosy,
cytomegalovirus (CMV), respiratory syncytial virus,
Epstein Barr virus, herpes, influenza, parainfluenza,
rotavirus, adenovirus, human immunodeficiency virus
(HIV), hepatitis A, NonA NonB hepatitis, varicella,
rabies, yellow fever, Japanese encephalitis, flavirus,
dengue, toxoplasmosis, coccidiomycosis, schistosomiasis,
and malaria immunogens.

69. The method of claim 67 where the following
conditions apply:

iv) the treatment and control groups differ as to the
presence, dosage, timing or composition of at least one
immunogen which is other than a live vaccine,

v) said immunogen is one other than a pertussis
immunogen.

70. The method of claim 67 where the following condition
applies:

xiii) at least one observation of the effect of the
schedules occurs at least five years after the birth of
the mammals.

71. The method of claim 67 where the following condition
applies:

x) at least one immunogen being studied for its effect on
the incidence or severity of said disorder is
administered starting after 41 days but before 180 days
of life.

72. The method of claim 71 where the following condition
applies:

xv) at least one immunogen administered to the treatment
group is selected from the group consisting of BCG,
measles, mumps, rubella, diphtheria, pertussis,
hemophilus influenza, tetanus, hepatitis B, polio,
anthrax, plague, encephalitis, meningococcal, meningitis,
pneumococcus, pneumonia, typhus, typhoid fever,

streptococcus, staphylococcus, neisseria, lyme disease,
cholera, E. coli, shigella, leishmania, leprosy,
cytomegalovirus (CMV), respiratory syncytial virus,
Epstein Barr virus, herpes, influenza, parainfluenza,
rotavirus, adenovirus, human immunodeficiency virus
(HIV), hepatitis A, NonA NonB hepatitis, varicella,
rabies, yellow fever, Japanese encephalitis, flavirus,
dengue, toxoplasmosis, coccidiomycosis, schistosomiasis,
and malaria immunogens

73.   The method of claim 66 where the following condition
applies:

x) at least one immunogen being studied for its effect on
the incidence or severity of said disorder is
administered starting after 41 days but before 180 days
of life.

74.   The method of claim 73 where the following condition
applies:

xv) at least one immunogen administered to the treatment
group is selected from the group consisting of BCG,
measles, mumps, rubella, diphtheria, pertussis,
hemophilus influenza, tetanus, hepatitis B, polio,
anthrax, plague, encephalitis, meningococcal, meningitis,
pneumococcus, pneumonia, typhus, typhoid fever,
streptococcus, staphylococcus, neisseria, lyme disease,
cholera, E. coli, shigella, leishmania, leprosy,
cytomegalovirus (CMV), respiratory syncytial virus,
Epstein Barr virus, herpes, influenza, parainfluenza,
rotavirus, adenovirus, human immunodeficiency virus
(HIV), hepatitis A, NonA NonB hepatitis, varicella,
rabies, yellow fever, Japanese encephalitis, flavirus,
dengue, toxoplasmosis, coccidiomycosis, schistosomiasis,
and malaria immunogens.

75.   The method of claim 73 where the following
conditions apply:

iv) the treatment and control groups differ as to the
presence, dosage, timing or composition of at least one
immunogen which is other than a live vaccine,

v) said immunogen is one other than a pertussis immunogen.

76.   The method of claim 73 where the following condition applies:

xiii) at least one observation of the effect of the schedules occurs at least five years after the birth of the mammals.

77.   The method of claim 1 where the following conditions apply:

i) the first dose of said immunization schedule is given in at least one group when the mammals are less than 42 days old,

ii) said immunization schedule prevents at least one infectious disease,

v) said immunogen is one other than a pertussis immunogen.

78.   The method of claim 77 where the following condition applies:

xviii) in at least one group both a pediatric immunogen and a non-pediatric immunogen are administered.

79.   The method of claim 1 where the following conditions apply:

i) the first dose of said immunization schedule is given in at least one group when the mammals are less than 42 days old,

ii) said immunization schedule prevents at least one infectious disease,

xvi) it is determined whether the age of the subject mammal, at the time of commencement of the immunization schedule, affects the incidence, prevalence or frequency of the disorder.

80. The method of claim 1 where the following conditions apply:

i) the first dose of said immunization schedule is given in at least one group when the mammals are less than 42 days old,

xviii) in at least one group both a pediatric immunogen and a non-pediatric immunogen are administered.

81. The method of claim 79 where the following condition applies:

xiii) at least one observation of the effect of the schedules occurs at least five years after the birth of the mammals.

82. The method of claim 80 where the following conditions apply:

ii) said immunization schedule prevents at least one infectious disease,

xiii) at least one observation of the effect of the schedules occurs at least five years after the birth of the mammals.

83. The method of claim 1 where the following conditions apply:

i) the first dose of said immunization schedule is given in at least one group when the mammals are less than 42 days old,

iv) the treatment and control groups differ as to the presence, dosage, timing or composition of at least one immunogen which is other than a live vaccine,

xiii) at least one observation of the effect of the schedules occurs at least five years after the birth of the mammals.

84. The method of claim 1 where the following conditions

apply:

> i) the first dose of said immunization schedule is given
> in at least one group when the mammals are less than 42
> days old,

> iv) the treatment and control groups differ as to the
> presence, dosage, timing or composition of at least one
> immunogen which is other than a live vaccine,

> xvi) it is determined whether the age of the subject
> mammal, at the time of commencement of the immunization
> schedule, affects the incidence, prevalence or frequency
> of the disorder.

85. The method of claim 1 where the following conditions
apply:

> i) the first dose of said immunization schedule is given
> in at least one group when the mammals are less than 42
> days old,

> iv) the treatment and control groups differ as to the
> presence, dosage, timing or composition of at least one
> immunogen which is other than a live vaccine,

> v) said immunogen is one other than a pertussis
> immunogen,

> xiii) at least one observation of the effect of the
> schedules occurs at least five years after the birth of
> the mammals.

86. The method of claim 1 where the following conditions
apply:

> i) the first dose of said immunization schedule is given
> in at least one group when the mammals are less than 42
> days old,

> x) at least one immunogen being studied for its effect on
> the incidence or severity of said disorder is
> administered starting after 41 days but before 180 days

of life,

ii) said immunization schedule prevents at least one
infectious disease,

v) said immunogen is one other than a pertussis
immunogen.

87. The method of claim 86 where the following condition
also applies:

xviii) in at least one group both a pediatric immunogen
and a non-pediatric immunogen are administered.

88. The method of claim 1 where the following conditions
apply:

x) at least one immunogen being studied for its effect on
the incidence or severity of said disorder is
administered starting after 41 days but before 180 days
of life,

ii) said immunization schedule prevents at least one
infectious disease,

v) said immunogen is one other than a pertussis
immunogen.

89. The method of claim 1 where the following conditions
apply:

x) at least one immunogen being studied for its effect on
the incidence or severity of said disorder is
administered starting after 41 days but before 180 days
of life,

iv) the treatment and control groups differ as to the
presence, dosage, timing or composition of at least one
immunogen which is other than a live vaccine,

xiii) at least one observation of the effect of the
schedules occurs at least five years after the birth of
the mammals.

90.   The method of claim 89 where the following condition also applies:

v) said immunogen is one other than a pertussis immunogen.

91.   The method of claim 1 where the following conditions apply:

x) at least one immunogen being studied for its effect on the incidence or severity of said disorder is administered starting after 41 days but before 180 days of life,

iv) the treatment and control groups differ as to the presence, dosage, timing or composition of at least one immunogen which is other than a live vaccine,

v) said immunogen is one other than a pertussis immunogen,

xvi) it is determined whether the age of the subject mammal, at the time of commencement of the immunization schedule, affects the incidence, prevalence or frequency of the disorder.

92.   The method of claim 91 where the following condition also applies:

vi) the method is part of a development process or clinical trial of a vaccine to test a vaccine for safety or efficacy.

93.   The method of claim 1 where the following conditions apply:

x) at least one immunogen being studied for its effect on the incidence or severity of said disorder is administered starting after 41 days but before 180 days of life,

iv) the treatment and control groups differ as to the presence, dosage, timing or composition of at least one

immunogen which is other than a live vaccine,

v) said immunogen is one other than a pertussis immunogen,

xvi) it is determined whether the age of the subject mammal, at the time of commencement of the immunization schedule, affects the incidence, prevalence or frequency of the disorder,

vi) the method is part of a development process or clinical trial of a vaccine to test a vaccine for safety or efficacy,

xv) at least one immunogen administered to the treatment group is selected from the group consisting of BCG, measles, mumps, rubella, diphtheria, pertussis, hemophilus influenza, tetanus, hepatitis B, polio, anthrax, plague, encephalitis, meningococcal, meningitis, pneumococcus, pneumonia, typhus, typhoid fever, streptococcus, staphylococcus, neisseria, lyme disease, cholera, E. coli, shigella, leishmania, leprosy, cytomegalovirus (CMV), respiratory syncytial virus, Epstein Barr virus, herpes, influenza, parainfluenza, rotavirus, adenovirus, human immunodeficiency virus (HIV), hepatitis A, NonA NonB hepatitis, varicella, rabies, yellow fever, Japanese encephalitis, flavirus, dengue, toxoplasmosis, coccidiomycosis, schistosomiasis, and malaria immunogens.

94. The method of claim 93 where the following condition also applies:

xiii) at least one observation of the effect of the schedules occurs at least five years after the birth of the mammals.

95. The method of claim 94 where the following condition also applies

i) the first dose of said immunization schedule is given in at least one group when the mammals are less than 42 days old.

96. The method of claim 1 where the following conditions apply:

    x) at least one immunogen being studied for its effect on
    the incidence or severity of said disorder is
    administered starting after 41 days but before 180 days
    of life,

    ii) said immunization schedule prevents at least one
    infectious disease,

    iv) the treatment and control groups differ as to the
    presence, dosage, timing or composition of at least one
    immunogen which is other than a live vaccine.

    97. A method of determining whether an immunization
schedule affects the incidence, prevalence or frequency of a
chronic immune-mediated disorder in a treatment group of
mammals, relative to a control group of mammals, which
comprises immunizing mammals in the treatment group of mammals
with one or more doses of one or more immunogens, according to
said immunization schedule, and comparing the incidence,
prevalence, or frequency of said chronic immune-mediated
disorder, in the treatment group, with that in the control
group,

wherein said chronic immune mediated disorder is a diabetes
mellitus which has not been chemically induced by
streptozotocin,

wherein said immunogen is one other than a BCG immunogen,
wherein the mammal is an animal model for human diabetes, and
where the control and the treatment group differ by at least
one of the following:

    (a) the presence of at least one immunogen,
    (b) the size of the dose of at least one immunogen,
    (c) the number of doses of at least one immunogen, or
    (d) the timing of administration of the first dose of at
         least one immunogen.

    98. The method of claim 97 in which

(i) the first dose is given to at least one group when the mammals are less than 42 days old,

99. The method of claim 97 where at least one of the following limitations applies:

i) the first dose is given to at least one group when the mammals are less than 42 days old,

ii) said immunization schedule protects against at least two infectious diseases if administered to humans,

iii) at least one immunogen is administered by a route other than intravenously,

iv) at least one immunogen is one other than a live vaccine,

v) said immunogen is one other than a pertussis immunogen,

vi) the method is part of a development process to test a vaccine for efficacy or safety,

vii) it is determined whether the age of the subject mammal, at the time of commencement of the immunization schedule, affects the incidence, prevalence or frequency of the disorder,

viii) at least one group receives more than one dose of at least one immunogen in a plurality of doses of said immunogen,

ix) at least one said treatment group receives one potentially pharmaceutically acceptable dose of each of at least two potentially pharmaceutically acceptable immunogenic agents, which comprise at least one potentially pharmaceutically acceptable first pediatric immunogen and at least one agent selected from the group consisting of a second pediatric immunogen and a nonpediatric immunogen,

x) at least one immunogen being studied for its effect

on the incidence, prevalence or frequency of said
disorder is administered starting after 41 days but
before 180 days of life,

xi) the method is part of a production process to test
vaccine lots for efficacy or safety,

xii) said mammals are NOD mice or BB rats,

xiii) the mammals are rodents and the groups are
compared from first administration until at least 24.5
weeks of age,

xiv) at least a second chronic immune-mediated disorder
in addition to diabetes is so compared, and said second
disorder is an autoimmune disease,

xv) at least one group receives an immunogen starting
after 41 days of life,

xvi) at least one immunogen is selected from the group
consisting of BCG, measles, mumps, rubella, diphtheria,
pertussis, hemophilus influenza, tetanus, hepatitis B,
polio, anthrax, plague, encephalitis, meningococcal,
meningitis, pneumococcus, pneumonia, typhus, typhoid
fever, streptococcus, staphylococcus, neisseria, lyme
disease, cholera, E. coli, shigella, leishmania, leprosy,
cytomegalovirus (CMV), respiratory syncytial virus,
Epstein Barr virus, herpes, influenza, parainfluenza,
rotavirus, adenovirus, human immunodeficiency virus
(HIV), hepatitis A, NonA NonB hepatitis, varicella,
rabies, yellow fever, Japanese encephalitis, flavirus,
dengue, toxoplasmosis, coccidiomycosis, schistosomiasis,
and malaria immunogens

xvii) said groups have not received an autoantigen
capable of inducing diabetes,

xviii) at least two pediatric immunogens are
administered to at least one group,

xvix) the mammals are made susceptible to said immune-
mediated disorder by administration of an

immunosuppressant or by immunosuppressive surgery,

xx) the mammals are rodents and the groups are compared from first administration until at least 52 days after the last administration of an immunogen according to the immunization schedules for said groups,

xxi) the level of an autoantibody marker in two groups is compared,

xxii) at least one immunogen is administered with a depot adjuvant,

xxiii) a dose of immunogen being administered is a pharmaceutically acceptable dose or a dose indicating that a pharmaceutically acceptable dose would reduce said incidence or severity of said disorder or the level of a marker of such a disorder,

xxiv) the mammals are rodents.

100. The method of claim 97 in which

vii the first dose is given when the mammals are less than 14 days old.

101. The method of claim 99 where at least 2 conditions apply.

102. The method of claim 99 where at least 3 conditions apply.

103. The method of claim 99 were at least 4 conditions apply.

104. The method of claim 99 were at least 5 conditions apply.

105. The method of claim 99 were at least 6 conditions apply.

106. The method of claim 99 were at least 7 conditions apply.

107. The method of claim 99 where at least 8 conditions apply.

108. The method of claim 99 where at least 9 conditions apply.

109. The method of claim 99 where at least 10 conditions apply.

110. The method of claim 99 where at least 11 conditions apply.

111. The method of claim 99 where at least 13 conditions apply.

112. The method of claim 99 where at least 14 conditions apply.

113. The method of claim 99 where at least 15 conditions apply.

114. The method of claim 99 were at least 16 conditions apply.

115. The method of claim 99 were at least 17 conditions apply.

116. The method of claim 99 were at least 18 conditions apply.

117. The method of claim 99 were at least 19 conditions apply.

118. The method of claim 99 where at least 20 conditions apply.

119. The method of claim 99 where at least 21 conditions apply.

120. The method of claim 99 where at least 22 conditions apply.

121. The method of claim 99 where at least 23 conditions

apply.

122. The method of claim 99 where at least 24 conditions apply.

123. The method of claim 97 where the following conditions apply:

i) the first dose is given to at least one group when the mammals are less than 42 days old,

ii) said immunization schedule involves administration of immunogens capable of protecting against at least one infectious disease in humans,

iv) at least one immunogen is one other than a live vaccine,

v) said immunogen is one other than a pertussis immunogen.

124. The method of claim 123 where the following condition applies:

viii) at least one group receives more than one dose of at least one immunogen in a plurality of doses of said immunogen.

125. The method of claim 123 where the following condition applies:

x) at least one immunogen being studied for its effect on the incidence, prevalence or frequency of said disorder is administered starting after 41 days but before 180 days of life.

126. The method of claim 123 where the following condition applies:

xxiii) a dose of immunogen being administered is a pharmaceutically acceptable dose or a dose indicating that a pharmaceutically acceptable dose would reduce said incidence or severity of said disorder or the level of a

marker of such a disorder,

127. The method of claim 123 where the following condition applies:

xxiv) the mammals are rodents.

128. The method of claim 127 where the following condition applies:

xii) said mammals are NOD mice or BB rats.

129. The method of claim 123 where the following condition applies:

xiii) the mammals are rodents and the groups are compared from first administration until at least 24.5 weeks of age.

130. The method of claim 123 where the following condition applies:

xx) the mammals are rodents and the groups are compared from first administration until at least 52 days after the last administration of an immunogen according to the immunization schedules for said groups.

131. The method of claim 123 where the following condition applies:

vi) the method is part of a development process to test a vaccine for efficacy or safety.

132. The method of claim 123 where the following condition applies:

xi) the method is part of a production process to test vaccine lots for efficacy or safety.

133. The method of claim 123 where the following condition applies:

xvi) at least one immunogen is selected from the group

consisting of BCG, measles, mumps, rubella, diphtheria,
pertussis, hemophilus influenza, tetanus, hepatitis B,
polio, anthrax, plague, encephalitis, meningococcal,
meningitis, pneumococcus, pneumonia, typhus, typhoid
fever, streptococcus, staphylococcus, neisseria, lyme
disease, cholera, E. coli, shigella, leishmania, leprosy,
chytomegalovirus (CMV), respiratory syncytial virus,
Epstein Barr virus, herpes, influenze, parainfluenze,
rotavirus, adenovirus, human immunodeficiency virus
(HIV), hepatitis A, NonA NonB hepatitis, varicella,
rabies, yellow fever, rabies, Japanese encephalitis,
flavirus, dengue toxoplasmosis, coccidiomycosis,
schistosomiasis, and malaria immunogens.

134. The method of claim 123 where the following
condition applies:

iii) at least one immunogen is administered by a route
other than intravenously.

135. The method of claim 123 where the following
conditions apply:

xiii) the mammals are rodents and the groups are
compared from first administration until at least 24.5
weeks of age,

xvi) at least one immunogen is selected from the group
consisting of BCG, measles, mumps, rubella, diphtheria,
pertussis, hemophilus influenza, tetanus, hepatitis B,
polio, anthrax, plague, encephalitis, meningococcal,
meningitis, pneumococcus, pneumonia, typhus, typhoid
fever, streptococcus, staphylococcus, neisseria, lyme
disease, cholera, E. coli, shigella, leishmania, leprosy,
chytomegalovirus (CMV), respiratory syncytial virus,
Epstein Barr virus, herpes, influenze, parainfluenze,
rotavirus, adenovirus, human immunodeficiency virus
(HIV), hepatitis A, NonA NonB hepatitis, varicella,
rabies, yellow fever, rabies, Japanese encephalitis,
flavirus, dengue toxoplasmosis, coccidiomycosis,
schistosomiasis, and malaria immunogens.

136. The method of claim 135 where the following

condition applies:

> viii) at least one group receives more than one dose of
> at least one immunogen in a plurality of doses of said
> immunogen.

137. The method of claim 135 where the following
condition applies:

> x) at least one immunogen being studied for its effect
> on the incidence, prevalence or frequency of said
> disorder is administered starting after 41 days but
> before 180 days of life.

138. The method of claim 135 where the following
condition applies:

> xii) said mammals are NOD mice or BB rats.

139. The method of claim 135 where the following
condition applies:

> xx) the mammals are rodents and the groups are compared
> from first administration until at least 52 days after
> the last administration of an immunogen according to the
> immunization schedules for said groups.

140. The method of claim 135 where the following
condition applies:

> xi) the method is part of a production process to test
> vaccine lots for efficacy or safety.

141. The method of claim 135 where the following
condition applies:

> iii) at least one immunogen is administered by a route
> other than intravenously.

142. The method of claim 135 where the following
condition applies:

> ix) at least one said treatment group receives one

potentially pharmaceutically acceptable dose of each of
at least two potentially pharmaceutically acceptable
immunogenic agents, which comprise at least one
potentially pharmaceutically acceptable first pediatric
immunogen and at least one agent selected from the group
consisting of a second pediatric immunogen and a
nonpediatric immunogen.

143. The method of claim 123 where the following
condition applies

ix) at least one said treatment group receives one
potentially pharmaceutically acceptable dose of each of
at least two potentially pharmaceutically acceptable
immunogenic agents, which comprise at least one
potentially pharmaceutically acceptable first pediatric
immunogen and at least one agent selected from the group
consisting of a second pediatric immunogen and a
nonpediatric immunogen.

144. The method of claim 125 where the following
condition applies

ix) at least one said treatment group receives one
potentially pharmaceutically acceptable dose of each of
at least two potentially pharmaceutically acceptable
immunogenic agents, which comprise at least one
potentially pharmaceutically acceptable first pediatric
immunogen and at least one agent selected from the group
consisting of a second pediatric immunogen and a
nonpediatric immunogen.

145. The method of claim 128 where the following
condition applies

ix) at least one said treatment group receives one
potentially pharmaceutically acceptable dose of each of
at least two potentially pharmaceutically acceptable
immunogenic agents, which comprise at least one
potentially pharmaceutically acceptable first pediatric
immunogen and at least one agent selected from the group
consisting of a second pediatric immunogen and a
nonpediatric immunogen.

146. The method of claim 137 where the following condition applies

ix) at least one said treatment group receives one potentially pharmaceutically acceptable dose of each of at least two potentially pharmaceutically acceptable immunogenic agents, which comprise at least one potentially pharmaceutically acceptable first pediatric immunogen and at least one agent selected from the group consisting of a second pediatric immunogen and a nonpediatric immunogen.

147. The method of claim 138 where the following condition applies

ix) at least one said treatment group receives one potentially pharmaceutically acceptable dose of each of at least two potentially pharmaceutically acceptable immunogenic agents, which comprise at least one potentially pharmaceutically acceptable first pediatric immunogen and at least one agent selected from the group consisting of a second pediatric immunogen and a nonpediatric immunogen.
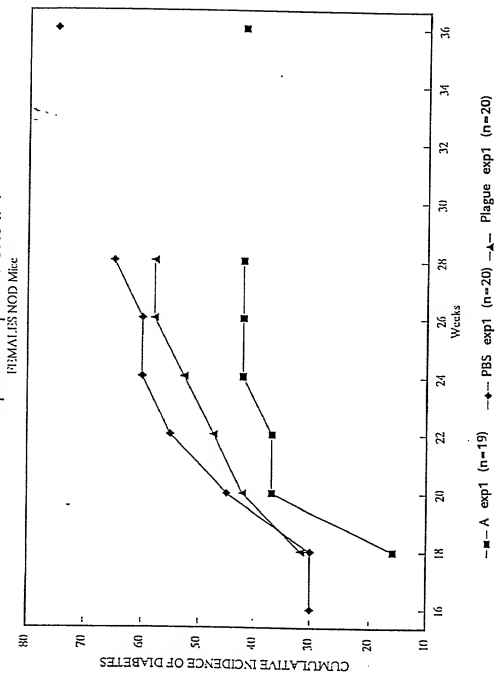
148. The method of claim 1 wherein at least one of said immunization schedules protects against at least one infectious disease.
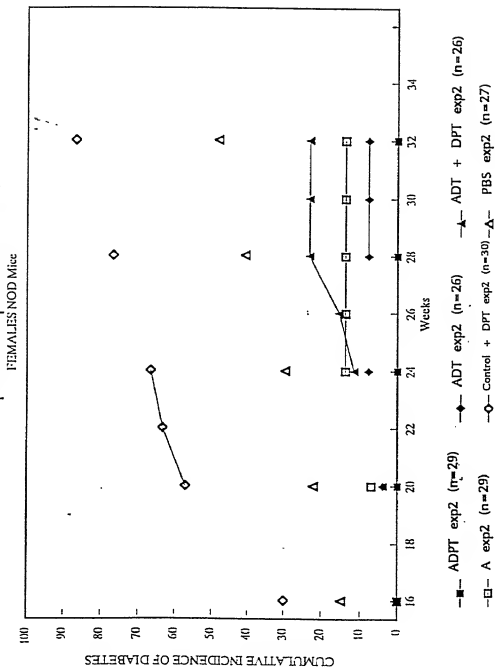
ABSTRACT

A method of immunization, and compositions therefor, are provided for substantially preventing or reducing the symptoms of at least one infectious disease and at least one chronic immune mediated disorder. An immunogenic challenge which supplements the normal childhood immunization schedule can help ensure the proper maturation of the immune system and prevent the development of chronic immune mediated disorders, such as immune-mediated diabetes or SLE.

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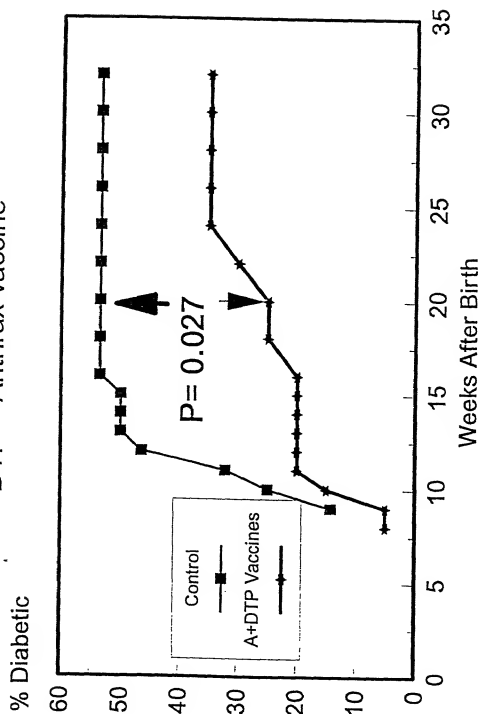
FIGURE 1

Graph I. Experiment #1

FEMALES NOD Mice

CUMULATIVE INCIDENCE OF DIABETES

Weeks

—■— A exp1 (n=19)   —◆— PBS exp1 (n=20)   —▲— Plague exp1 (n=20)

FIGURE 2

Graph II. Experiment #2

FEMALES' NOD Mice

CUMULATIVE INCIDENCE OF DIABETES

ADPT exp2 (n=29)

A exp2 (n=29)

ADT exp2 (n=26)

Control + DPT exp2 (n=30)

ADT + DPT exp2 (n=26)

PBS exp2 (n=27)

Figure 3
BB Rats
DTP + Anthrax vaccine

% Diabetic

P= 0.027

Control
A+DTP Vaccines

Weeks After Birth

Page 1 of 1   [X] Original   [ ] Supplemental   Atty.Docket: CLASSEN = 1A

Combined Declaration for Patent Application and Power of Attorney

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name; and that I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled METHOD AND COMPOSITION FOR AN EARLY VACCINE TO PROTECT AGAINST BOTH COMMON INFECTIOUS DISEASES AND CHRONIC IMMUNE MEDIATED DISORDERS OR THEIR SEQUELAE
the specification of which (check one)

    [ ] is attached hereto;
    [ ] was filed in the United States under 35 U.S.C. §111 on _____, as USSN _____*; or
    [XX] was/will be filed in the U.S. under 35 U.S.C. §371 by entry into the U.S. national stage of an international (PCT)
        application, PCT/US94/08825_____, filed 04 August 1994 _____, entry requested
        on _____*; national stage application received USSN _____*:
        §371/§102(e) date _____* (*if known),
and was amended on 10 October 1995 _____ (if applicable)

(include dates of amendments under PCT Art. 19 and 34 if PCT)

I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above; and I acknowledge the duty to disclose to the Patent and Trademark Office (PTO) all information known by me to be material to patentability as defined in 37 C.F.R. § 1.56.

I hereby claim foreign priority benefits under 35 U.S.C. §§ 119, 365 of any prior foreign application(s) for patent or inventor's certificate, or prior PCT application(s) designating a country other than the U.S., listed below with the "Yes" box checked and have also identified below any such application having a filing date before that of the application on which priority is claimed:

(Number)	(Country)	(Day Month Year Filed)	[ ] YES	[ ] NO
(Number)	(Country)	(Day Month Year Filed)	[ ] YES	[ ] NO
(Number)	(Country)	(Day Month Year Filed)	[ ] YES	[ ] NO

I hereby claim the benefit under 35 U.S.C. § 120 of any prior U.S. Application(s) or prior PCT Application(s) designating the U.S. listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in such U.S. or PCT application in the manner provided by the first paragraph of 35 U.S.C. §112, I acknowledge the duty to disclose to the PTO all information as defined in 37 C.F.R. §1.56(a) which occurred between the filing date of the prior application and the national filing date of this application:

08/104,529	12 August 1993	pending
(Application Serial No.)	(Day Month Year Filed)	(Status: patented, pending, abandoned)
(Application Serial No.)	(Day Month Year Filed)	(Status: patented, pending, abandoned)

I hereby appoint the following attorneys, with full power of substitution, association, and revocation, to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith.

  SHERIDAN NEIMARK, REG. NO. 20,520 • ROGER L. BROWDY, REG. NO. 25,618 ANNE H. KINNAMAN, REG. NO. 25,894
  NORMAN J. LATKER, REG. NO. 19,963 • IVER P. COOPER, REG. NO. 28,805 • A. FRED STANGHEIN, REG. NO. 18,453
  ALLEN C. YUN, REG. NO. 37,971*   NICK K. GROENER, REG. NO. 33,478* * Patent Agent

ADDRESS ALL CORRESPONDENCE TO **BROWDY AND NEIMARK** 419 Seventh Street, N.W. Washington, D.C. 20004	DIRECT ALL TELEPHONE CALLS TO: **BROWDY AND NEIMARK** (202) 628-5197

I hereby further declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under 18 U.S.C. §1001 and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

FULL NAME OF SOLE OR FIRST INVENTOR John Barthelow Classen	INVENTOR'S SIGNATURE John B Classen	DATE 2/9/96
RESIDENCE Baltimore, Maryland		CITIZENSHIP American
POST OFFICE ADDRESS 6517 Montrose Avenue, Baltimore, Maryland 21212, United States of America		
FULL NAME OF SECOND JOINT INVENTOR	INVENTOR'S SIGNATURE	DATE
RESIDENCE		CITIZENSHIP
POST OFFICE ADDRESS		
FULL NAME OF THIRD JOINT INVENTOR	INVENTOR'S SIGNATURE	DATE
RESIDENCE		CITIZENSHIP
POST OFFICE ADDRESS		

ALL INVENTORS MUST REVIEW APPLICATION AND DECLARATION BEFORE SIGNING. ALL ALTERATIONS MUST BE INITIALED AND DATED BY ALL INVENTORS PRIOR TO EXECUTION. NO ALTERATIONS CAN BE MADE AFTER THE DECLARATION IS SIGNED. ALL PAGES OF DECLARATION MUST BE SEEN BY ALL INVENTORS.

applicant or Patentee: _John Classen_   Attorney's Docket No.: _CLASSEN=1A_
Serial or Patent No.:   Filed or Issued:
For: _METHOD AND COMPOSITION FOR AN EARLY VACCINE TO PROTECT AGAINST BOTH COMMON INFECTIOUS DISEASES AND..._

VERIFIED STATEMENT (DECLARATION) CLAIMING SMALL BUSINESS ENTITY STATUS
(37 CFR 1.9(f) and 1.27(C)) - SMALL BUSINESS CONCERN

I hereby declare that I am
[X] the owner of the small business concern identified below:
[ ] an official of the small business concern empowered to act on behalf of the concern identified below:

NAME OF CONCERN   CLASSEN IMMUNOTHERAPIES, INC.
ADDRESS OF CONCERN   6517 Montrose Avenue, Baltimore, Maryland 21212, United States of America

I hereby declare that the above identified small business concern qualifies as a small business concern as defined in 13 CFR 121.3-18, and reproduced in 37 CFR 1.9(d), for purposes of paying reduced fees under section 41(a) and (b) of Title 35, United States Code, in that the number of employees of the concern, including those of its affiliates, does not exceed 500 persons. For purposes of this statement, (1) the number of employees of the business concern is the average over the previous fiscal year of the concern of the persons employed on a full-time, part-time or temporary basis during each of the pay periods of the fiscal year, and (2) concerns are affiliates of each other when either, directly or indirectly, one concern controls or has the power to control the other, or a third party or parties controls or has the power to control both.

I hereby declare that rights under contract or law have been conveyed to and remain with the small business concern identified above with regard to the invention entitled _METHOD AND COMPOSITION FOR AN EARLY VACCINE TO PROTECT AGAINST BOTH COMMON INFECTIOUS DISEASES AND CHRONIC IMMUNE MEDIATED DISORDERS OR THEIR SEQUELAE_ by inventor _John CLASSEN_
described in:

  [ ] the specification filed herewith
  [XX] application serial no. _PCT/US94/08825_____, filed _04 August 1994_____.
  [ ] patent no. _____, issued _____.

If the rights held by the above-identified small business concern are not exclusive, each individual, concern or organization having rights to the invention is listed below* and no rights to the invention are held by any person, other than the inventor, who would not qualify as an independent inventor under 37 CFR 1.9(c) if that person made the invention, or by any concern which would not qualify as a small business concern under 37 CFR 1.9(d), or a nonprofit organization under 37 CFR 1.9(e).

          *Note: Separate verified statements are required from each named person, concern or organization having rights
          to the invention averring to their status as small entities (37 CFR 1.27).

NAME _____
ADDRESS _____
          [ ] INDIVIDUAL   [ ] SMALL BUSINESS CONCERN   [ ] NONPROFIT ORGANIZATION
NAME _____
ADDRESS _____
          [ ] INDIVIDUAL   [ ] SMALL BUSINESS CONCERN   [ ] NONPROFIT ORGANIZATION

I acknowledge the duty to file, in this application or patent, notification of any change in status resulting in loss of entitlement to small entity status prior to paying, or at the time of paying, the earliest of the issue fee or any maintenance fee due after the date on which status as a small entity is no longer appropriate. (37 CFR 1.28(b))
I hereby declare that all statements made herein of my own knowledge are true and that all statements on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application, any patent issuing thereon, or any patent to which this verified statement is directed.

NAME OF PERSON SIGNING   _John B C(lassen_   (JOHN B   CLASSEN)
TITLE OF PERSON SIGNING OTHER THAN OWNER   _OWNER_
ADDRESS OF PERSON SIGNING   _6517 MONTROSE   AVE, G.LTO, MD   21212_
SIGNATURE _John B Classen_   DATE _2/9/96_